

## Combined effects of CO<sub>2</sub> and temperature on carbon uptake and partitioning by the marine diatoms *Thalassiosira weissflogii* and *Dactyliosolen fragilissimus*

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### Abstract

Carbon uptake and partitioning of two globally abundant diatom species, *Thalassiosira weissflogii* and *Dactyliosolen fragilissimus*, was investigated in batch culture experiments under four conditions: ambient (15°C, 400 μatm), high CO<sub>2</sub> (15°C, 1000 μatm), high temperature (20°C, 400 μatm), and combined (20°C, 1000 μatm). The experiments were run from exponential growth into the stationary phase (six days after nitrogen depletion), allowing us to track biogeochemical dynamics analogous to bloom situations in the ocean. Elevated CO<sub>2</sub> had a fertilizing effect and enhanced uptake of dissolved inorganic carbon (DIC) by about 8% for *T. weissflogii* and by up to 39% for *D. fragilissimus*. This was also reflected in higher cell numbers, build-up of particulate and dissolved organic matter, and transparent exopolymer particles. The CO<sub>2</sub> effects were most prominent in the stationary phase when nitrogen was depleted and CO<sub>2</sub>(aq) concentrations were low. This indicates that diatoms in the high CO<sub>2</sub> treatments could take up more DIC until CO<sub>2</sub> concentrations in seawater became so low that carbon limitation occurs. These results suggest that, contrary to common assumptions, diatoms could be highly sensitive to ongoing changes in oceanic carbonate chemistry, particularly under nutrient limitation. Warming from 15 to 20 °C had a stimulating effect on one species but acted as a stressor on the other species, highlighting the importance of species-specific physiological optima and temperature ranges in the response to ocean warming. Overall, these sensitivities to CO<sub>2</sub> and temperature could have profound impacts on diatoms blooms and the biological pump.

Over the past few centuries, atmospheric carbon dioxide (CO<sub>2</sub>) concentrations increased from preindustrial levels of approximately 280 ppmv (parts per million volume) to around 400 ppmv in the year 2013 (IPCC 2007). This increase in atmospheric CO<sub>2</sub> leads to a global increase in temperatures and its subsequent uptake by the ocean results in a decrease in seawater pH, both occurring at a pace unprecedented in recent geological history (IPCC 2007; Doney et al. 2009). The reduction in seawater pH, a phenomenon commonly referred to as ocean acidification (Caldeira and Wickett 2003), is accompanied by changes in carbonate chemistry, leading to a strong decrease in carbonate ion (CO<sub>3</sub><sup>2-</sup>) concentration, a slight increase in bicarbonate (HCO<sub>3</sub><sup>-</sup>), and a strong increase in dissolved CO<sub>2</sub> and protons (H<sup>+</sup>) (Zeebe and Wolf-Gladrow 2001). Both, changes in CO<sub>2</sub> and temperature affect the buffering capacity of seawater, which is usually given as the fractional change in CO<sub>2</sub> over the fractional change in dissolved inorganic carbon

(DIC) (Revelle factor):  $(\Delta\text{CO}_2/\text{CO}_2)/(\Delta\text{DIC}/\text{DIC})$  (Egleston et al. 2010). At the same time, global average sea surface temperatures have already increased by 0.6°C since the beginning of the 20th century. Recent climate projections suggest an increase in global surface air temperatures of 1.1–6.4°C by the end of this century (relative to 1980–1999), thereby also leading to a further warming of the upper ocean (IPCC 2007). Both warming and ocean acidification are expected to affect marine ecosystems and carbon cycling in a variety of ways (for reviews see, e.g., Riebesell et al. 2009, Passow and Carlson 2012).

Temperature is a major environmental driver controlling metabolic rates, such as enzymatic reactions, thereby substantially affecting phytoplankton physiology (Duarte 2007; Boyd et al. 2013). Furthermore, it has been found that ocean warming might lead to a decrease in cell size (Moran et al. 2010), shifts in phenology (Sommer and Lengfellner 2008), and alterations in carbon cycling by phytoplankton (Wohlers et al. 2009; Taucher et al. 2012). Concurrently, ocean acidification is expected to influence marine phytoplankton

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either through a decrease in pH (and the corresponding increase in H<sup>+</sup> ions) or the shift in carbonate speciation, e.g., the relative availability of CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> for carbon assimilation and photosynthesis. Over the last few years, an increasing number of studies investigated effects of ocean acidification on various functional groups of phytoplankton, showing, for example, a marked influence of carbonate chemistry on growth and calcification in coccolithophores (Riebesell et al. 2000; Langer et al. 2006; Iglesias-Rodriguez et al. 2008) and nitrogen fixation in cyanobacteria (Hutchins et al. 2009). Furthermore, it has been demonstrated that simultaneous changes in temperature and CO<sub>2</sub> can have interactive effects on growth rates, photosynthesis, and elemental composition of various marine phytoplankton (Feng et al. 2008; Fu et al. 2007, 2008).

Relatively few studies exist about the effects of ocean acidification on diatoms, which account for 40% of total primary production in the ocean (Nelson et al. 1995), frequently form massive blooms, and are commonly assumed to dominate export production (Lampitt 1985; Treguer et al. 1995; Buesseler 1998), making them major players in marine biogeochemical cycling and sequestration of carbon to the deep ocean (Sarhou et al. 2005; Boyd et al. 2010).

Diatoms, like most other noncalcifying phytoplankton, are assumed to be affected by ocean acidification primarily through changes in the availability of dissolved CO<sub>2</sub> as a substrate for photosynthesis (Rost et al. 2008). Concentrations of CO<sub>2</sub> in the surface ocean are relatively low compared to other forms of inorganic carbon, especially HCO<sub>3</sub><sup>-</sup>. Yet, the primary carboxylating enzyme used in photosynthesis, RubisCO, is restricted to CO<sub>2</sub> for fixation of inorganic carbon, even though its substrate affinity for CO<sub>2</sub> is rather low. Therefore, in addition to passive uptake of CO<sub>2</sub> via diffusion, most phytoplankton operate a carbon concentrating mechanism (CCM) to enhance their intracellular CO<sub>2</sub> relative to external concentrations (for a recent review see Reinfelder 2011). Although the exact functioning of the CCM varies greatly among and within the dominant groups of marine phytoplankton, it generally involves active uptake of CO<sub>2</sub> and/or HCO<sub>3</sub><sup>-</sup> into the cell interior, as well as external or intracellular activity of the enzyme carbonic anhydrase, which accelerates the otherwise slow conversion between HCO<sub>3</sub><sup>-</sup> and CO<sub>2</sub>. For diatoms, there is evidence that their CCM is highly regulated and efficient with respect to changes in CO<sub>2</sub> (Rost et al. 2003; Trimborn et al. 2009). Correspondingly, most studies find little or no change in photosynthetic rates of diatoms under elevated pCO<sub>2</sub>, suggesting that carbon assimilation is essentially saturated under present-day CO<sub>2</sub> levels and thus unlikely to be affected by ocean acidification (Burkhardt et al. 1999; Rost et al. 2008; Beardall et al. 2009). However, this is in sharp contrast to the observed effect of elevated CO<sub>2</sub> on inorganic carbon uptake by diatoms in natural plankton communities (Riebesell et al. 2007; Tortell et al. 2008).

This controversy might result from the fact that laboratory-based studies usually do not take into account bloom dynamics and the postbloom physiological states of diatoms. However, exponential growth under nutrient-replete conditions and constant carbonate chemistry are not necessarily reflective of behavior during a natural bloom, where nutrient limitation and substantial shifts in the carbonate system occur. Considering the energetic and nutrient costs associated with phytoplankton CCMs, it seems likely that potential effects of environmental factors such as temperature and CO<sub>2</sub> on the physiological performance of phytoplankton become more pronounced in the stationary phase when nutrients are depleted and resource allocation becomes critical (Giordano et al. 2005; Hopkinson et al. 2011). Furthermore, it has long been recognized that a substantial portion of carbon uptake can occur after nutrient depletion and lead to a notable decoupling of carbon and nitrogen uptake, a phenomenon commonly referred to as “carbon overconsumption” (Sambrotto et al. 1993; Toggweiler 1993). These dynamics during the declining phase of a bloom are particularly important for export production (Lampitt 1985) and results from previous mesocosm experiments with natural, diatom-dominated plankton communities indicate that carbon overconsumption is affected by both temperature (Taucher et al. 2012) and CO<sub>2</sub> (Riebesell et al. 2007). However, carbon uptake and its allocation between the particulate and dissolved phase by primary producers on the one side, and heterotrophic consumption of this organic matter on the other side, operate simultaneously and are closely intertwined under natural or seminatural conditions in such studies, making it difficult to disentangle the underlying mechanisms.

The aim of this study is to help resolve this controversy and specifically investigate the effects of warming and increased pCO<sub>2</sub> on carbon uptake by two marine diatom species, *Thalassiosira weissflogii* and *Dactyliosolen fragilissimus*, with particular focus on dynamics after N-depletion, as well as the partitioning of this carbon between the particulate and dissolved phase.

## Methods

### Pre-experiments to determine the temperature range of the diatoms

Pre-experiments (referred to as temperature gradient experiment throughout) were carried out to identify the temperature range and optimum of two diatom species, by setting up a temperature gradient from 10°C to 22.5°C.

The aim was to choose two species that co-occur in nature but have different temperature optima, so that warming in the main experiment had a stimulating effect on one species, but acted as a stressor on the other species, pushing it beyond its temperature optimum. Thereby, a range of potential responses to future ocean conditions, i.e., warming in

combination with increased CO<sub>2</sub>, and possible synergistic effects could be investigated in the main experiments. Ideally, we would have carried out the main experiments with temperatures below and above the optimum for both species. However, this was not feasible due to logistical reasons.

To identify temperature range and optimum of the diatom species, cultures were grown in gas-tight polycarbonate bottles (one liter) at ambient  $p\text{CO}_2$  (400  $\mu\text{atm}$ ) under six temperatures (10.0°C, 12.5°C, 15.0°C, 17.5°C, 20.0°C, and 22.5°C) that were established with the help of temperature controlled water baths in two climate-controlled rooms. Prior to incubation at target temperatures, both diatom species were preacclimated to 12.5°C and 20°C for the lower and upper three temperature levels, respectively, for one week. Light supply and nutrient availability were identical to those of the main experiment (see below). Daily measurements were restricted to pH, NO<sub>3</sub><sup>-</sup>, and in vivo fluorescence ( $F_T$ ) due to the small volume of the incubation bottles (see Sampling and measurements section for details on measurement methods). Additional samples for particulate and dissolved organic matter (DOM) were taken on the first day of the experiment ( $t_0$ ), the day the respective cultures reached NO<sub>3</sub> depletion ( $t_{\text{NO}}$ ), and four days after NO<sub>3</sub> depletion ( $t_{\text{N4}}$ ). Optimum temperature was defined as that supporting maximal growth rates as determined by daily changes in fluorescence.

## Experimental setup

### Phytoplankton strains

All experiments presented here were conducted with monoclonal cell cultures of two globally abundant diatom species (Leblanc et al. 2012), using the single-celled and rather small species *T. weissflogii* (10–20  $\mu\text{m}$  cell diameter, strain CCMP 1336), and the larger, chain-forming species *D. fragilissimus* (50–100  $\mu\text{m}$  cell diameter, isolated from the North Sea in September 2012, now listed as strain RCC3389).

### Treatments: Combination of temperature and CO<sub>2</sub>

For each diatom species, eight gas-tight polyethylene bags were set up in duplicate in a combination of two temperatures and two  $p\text{CO}_2$  levels, with temperatures of 15°C and 20°C, initial partial pressure levels of CO<sub>2</sub> of 400  $\mu\text{atm}$  and 1000  $\mu\text{atm}$ . In the following, these treatments will be referred to as: “ambient” (15°C and 400  $\mu\text{atm}$ ), “high CO<sub>2</sub>” (15°C and 1000  $\mu\text{atm}$ ), “warming” (20°C and 400  $\mu\text{atm}$ ), and “combined” (20°C and 1000  $\mu\text{atm}$ ), respectively. Light supply was set to a photon flux density of  $\sim 100 \mu\text{mol m}^{-2} \text{s}^{-1}$  and a light/dark cycle of 14/10 h in all treatments.

### Materials, seawater, and nutrients

Collapsible bags (100 × 40 cm) constructed out of six millimeter polyethylene were used as incubation vessels for these experiments and allowed us to incubate the cultures without the presence of a headspace. The bags were sus-

pending in large water baths to maintain temperature and minimize gas exchange through the bag walls. The initial volume of the bags was 20 L. As volume was removed from the incubation the bags were rolled up to minimize the surface area to volume ratio. The bags were sterilized and dissolved organic carbon (DOC) leaching minimized by soaking in 10% HCl for at least 48 h prior to the experiment. Preliminary experiments showed no significant change in DIC or DOC over a period of 10 d in cleaned bags containing only sterile artificial seawater (ASW).

The experiment with *T. weissflogii* was conducted with ASW, following Kester et al. (1967). Since, *D. fragilissimus* did not grow well in ASW, experiments with this species were carried out in natural seawater (NSW) from the Santa Barbara Channel, which was collected in late April 2013 and subsequently filtered through 1.2  $\mu\text{m}$  and 0.2  $\mu\text{m}$  filters. Media was not autoclaved to avoid perturbation of the carbonate chemistry beyond the range found in NSW. Both culture media (ASW and NSW) were instead treated with a UV lamp (50 Watt) for 30 min before filling into the bags to minimize DOC and bacterial contamination. Preliminary experiments showed that such a UV-treatment reduced bacterial production (BP) below detection limit for at least a week.

ASW and NSW were enriched with inorganic nutrients in a modified version of f/2 medium (Guillard and Ryther 1962), with concentrations calculated in such a way that the diatom cultures became nitrate limited. Initial concentrations of NO<sub>3</sub><sup>-</sup> were identical in both experiments, amounting to 15  $\mu\text{mol L}^{-1}$ . Starting concentrations for PO<sub>4</sub><sup>3-</sup> were 6  $\mu\text{mol L}^{-1}$  and 8  $\mu\text{mol L}^{-1}$ , and 16  $\mu\text{mol L}^{-1}$  and 50  $\mu\text{mol L}^{-1}$  for Si(OH)<sub>4</sub> in the experiments with *T. weissflogii* and *D. fragilissimus*, respectively. Trace metals and vitamins were added at f/8 levels. The higher concentrations of PO<sub>4</sub><sup>3-</sup> and Si(OH)<sub>4</sub> in the experiment with *D. fragilissimus* were based on preliminary experiments, which suggested a possibly higher Si and P demand compared to *T. weissflogii*. Furthermore, the ASW was further enriched with 2 ml L<sup>-1</sup> of filtered (0.2  $\mu\text{m}$ ) and UV-blasted NSW to provide possible micronutrients that are not included in the recipe of Guillard and Ryther (1962).

### Carbonate chemistry manipulation

Carbonate chemistry in our experiments was manipulated following the “closed system approach”, i.e., adjusting  $p\text{CO}_2$  at constant total alkalinity (TA) through combined addition of inorganic carbon, and acid or base to an incubation container without headspace (Rost et al. 2008). Calculations for the carbonate system manipulations were carried out with the program CO2SYS (Lewis and Wallace 1998), using the dissociation constants for carbonic acid as refitted by Dickson and Millero (1987) and taking into account nutrient additions for TA adjustment as well as respective temperatures for target  $p\text{CO}_2$  levels.

For the experiments with ASW, the carbonate system was adjusted to target levels of  $p\text{CO}_2$  and TA of  $2350 \mu\text{mol kg}^{-1}$  through additions of 1 M solutions of  $\text{HCO}_3^-$  as DIC and NaOH/HCl to keep TA constant.

The carbonate system in the experiments with NSW had to be adjusted differently. The collected water had a pH of 7.76 and TA of  $\sim 2290 \mu\text{mol kg}^{-1}$ . From that, we calculated the necessary amount of  $\text{HCO}_3^-$  addition to set the desired  $p\text{CO}_2$  levels of  $400 \mu\text{atm}$  and  $1000 \mu\text{atm}$  for the respective temperatures ( $15^\circ\text{C}$  and  $20^\circ\text{C}$ ). To simulate the carbonate system dynamics of a natural plankton bloom,  $p\text{CO}_2$  was adjusted only initially, and was then allowed to change freely, as driven by biological dynamics. Measurements confirmed that the desired initial experimental conditions were within  $\sim 0.02$  pH units (or  $\sim 20 \mu\text{atm } p\text{CO}_2$ ) of target conditions.

#### Acclimatization phase

Prior to the experiments, the diatom cultures were acclimatized to the respective target conditions of temperature and  $p\text{CO}_2$  for at least a week ( $\sim 5$ – $6$  generations). During this acclimatization phase, the cultures were grown semicontinuously in gas-tight polycarbonate bottles (one liter). When  $p\text{CO}_2$  dropped below  $200 \mu\text{atm}$  (for the  $400 \mu\text{atm}$  treatment) or  $700 \mu\text{atm}$  (for the  $1000 \mu\text{atm}$  treatment), the cultures were diluted with fresh medium and  $p\text{CO}_2$  was readjusted to target conditions by combined additions of  $\text{HCO}_3^-$  and HCl or NaOH. After acclimatization, diatom cultures were inoculated at a density of  $500 \text{ cells mL}^{-1}$  (*T. weissflogii*) and  $200 \text{ cells mL}^{-1}$  (*D. fragillissimus*) to the respective experimental treatments.

#### Sampling and measurements

Sampling was carried out daily, immediately at the end of each light cycle. This maximized the comparability of samples between treatments and replicates, as all the cultures were sampled in the same physiological state. The bags were mixed gently prior to sampling to obtain representative water samples. Samples for pH and DOC were taken directly from the bag (gravity-filtration through in-line GF/F filter). For all other parameters, subsamples were collected in one liter polycarbonate bottles and later subdivided for various analyses. Since a major focus of our culture experiments was to determine carbon overconsumption and dynamics in the stationary phase, we continued sampling for (at least) six days after  $\text{NO}_3$  depletion.

pH was measured spectrophotometrically (Thermo Scientific Genesys 105 VIS Spectrophotometer with a SPG 1A air-cooled single cell Peltier element) at  $25^\circ\text{C}$  following Dickson et al. (2007), using *m*-cresol as an indicator dye. For maximum precision and accuracy, the dye was calibrated against certified reference material (A. Dickson, La Jolla, California).

DIC was measured on a nondispersive infrared analyzer following Bandstra et al. (2006). Samples were filtered through GF/F filters and stored in  $\sim 400 \text{ mL}$  borosilicate bot-

tles, sealed gas-tight, and stored at  $\sim 5^\circ\text{C}$  until analysis. Additional carbonate system variables were calculated from measured DIC and pH with the program CO2SYS (Lewis and Wallace 1998), using the dissociation constants for carbonic acid as refitted by Dickson and Millero (1987).

Samples for particulate organic carbon and nitrogen (POC and PON) were filtered onto precombusted (five hours at  $450^\circ\text{C}$ ) GF/F filters, rinsed with ASW immediately after filtering of samples in order to avoid accumulation of DOM on the filters, dried at  $\sim 60^\circ\text{C}$  for approximately 24 h and stored under vacuum in a desiccator until analysis. Afterward, they were analyzed on a CHN organic elemental analyzer (model CEC 440HA, Control Equipment Corp., now Exeter Analytical).

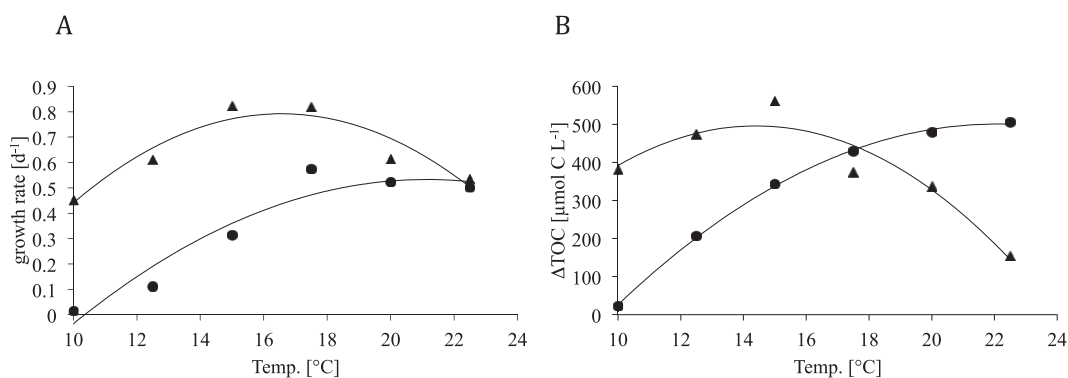
Samples for DOC were gravity-filtered through precombusted GF/F filters, with the filtrate collected in combusted (five hours at  $450^\circ\text{C}$ ) glass vials and frozen (at  $-20^\circ\text{C}$ ) until analysis. DOC was quantified via high-temperature combustion on a Shimadzu TOC-V as in Carlson et al. (2010), but with glucose calibration standards and Santa Barbara Channel reference standards that were routinely verified against DOM consensus reference water (Hansell 2005), for a resolution of  $1.5 \mu\text{mol L}^{-1}$ . Total dissolved nitrogen (TDN) was measured using the TOC-V (Farmer and Hansell 2007). Dissolved organic nitrogen (DON) was calculated by subtracting dissolved inorganic nitrogen (measured as described below) from TDN.

Samples for transparent exopolymer particles (TEP) were filtered onto  $0.4 \mu\text{m}$  polycarbonate filters and subsequently stained with Alcian Blue following the procedure of Passow and Alldredge (1995). TEP concentration was measured colorimetrically on a spectrophotometer (Thermo Scientific Genesys 105 VIS Spectrophotometer) through absorption at  $787 \text{ nm}$ . The dye solution had previously been calibrated using Gum Xanthan and the concentration of TEP is expressed as  $\mu\text{g}$  Gum Xanthan equivalents per liter ( $\mu\text{g GXeq L}^{-1}$ ). Conversion of TEP to carbon was not applied here, as conversion factors are highly species-specific and not available for *D. fragillissimus* to date (Engel and Passow 2001).

Samples for dissolved inorganic nitrate, nitrite, phosphate, and silicate were sterile-filtered through  $0.2 \mu\text{m}$  surfactant-free cellulose acetate filters, collected in 20-mL HDPE vials, and stored at  $-20^\circ\text{C}$  until analysis. Measurement was carried out on a flow injection analyzer (QuikChem 8000, Lachat Instruments, Zellweger Analytics).

BP was determined by the  $^3\text{H}$ -leucine incorporation ( $^3\text{H}$ -leu, SA =  $54.1 \text{ Ci mmo}^{-1}$ , Perkin Elmer, Boston, Massachusetts) method (Kirchman et al. 1986), using the microcentrifuge method (Smith and Azam 1992), modified by decanting supernatant rather than aspirating. Samples were taken on the first day of the experiment, as well as one and six days after N-depletion. The samples were then dark-incubated for 1.5–3 h and radioactivity was analyzed on a Beckman LS 6500 Liquid Scintillation Counter. All incubations were





**Fig. 1.** Effect of temperature on diatom growth and carbon cycling. Shown are (A) growth rate (d<sup>-1</sup>) under nutrient-replete conditions (calculated from fluorescence), (B) cumulative build-up of total organic carbon (μmol C L<sup>-1</sup>) from onset of the experiment until day 4 after N-depletion for *T. weissflogii* (circles) and *D. fragilissimus* (triangles). Solid lines denote significant relationship of temperature and the respective parameter detected by polynomial regression ( $p < 0.05$ ,  $n = 6$ ).

carried out in triplicate. To convert the incorporation of <sup>3</sup>H-leucine into carbon production (μg C L<sup>-1</sup> h<sup>-1</sup>), a theoretical conversion factor of 1.5 kg C mol<sup>-1</sup> leucine was used (Simon and Azam 1989).

Samples for cell concentrations were fixed with buffered formalin (40%) at a final concentration of 1% and counted microscopically using a hemocytometer.

In vivo dark-adapted chlorophyll fluorescence ( $F_T$ ) was measured using an AquaPen-C fluorometer (Photon Systems Instruments, Czech Republic). Samples were kept in the dark for at least 10 min before the measurement.

### Statistical analysis

For the temperature gradient experiment, a polynomial regression was fitted to the data to identify the temperature optimum of the two diatoms species ( $n = 6$ ). The analysis was based on growth rate as calculated from measured fluorescence during nutrient-replete growth as a function of temperature.

In the main experiments, we applied a fully crossed two-factorial design with temperature and  $p\text{CO}_2$  as the fixed factors and two treatment levels, respectively. Each treatment was set up in duplicates ( $n = 2$ ). All results below are given as the average value of replicates  $\pm$  standard error, which corresponds to the range of replicates for  $n = 2$ .

Statistical analysis for the detection of treatment effects was carried out with a permutational analysis of variance (PERMANOVA), because assumptions of normality and homogeneity of variances were not met for all analyzed data (Anderson 2001). For the analysis, “Euclidean distance” matrices with 9999 permutations were used, and pairwise (post hoc) tests were performed in case a significant effect ( $p < 0.05$ ) was detected. PERMANOVA was applied for maximum values of net DIC uptake and net build-up of POC and DOC, as well as their respective C : N ratios, relative to day 0 for each replicate. For cell concentrations and TEP, the aver-

age concentration in the stationary phase after nitrogen depletion was used, as measurements were highly variable in the experiment with *D. fragilissimus* after N-depletion.

### Results

#### Temperature range and optimum of the two diatom species

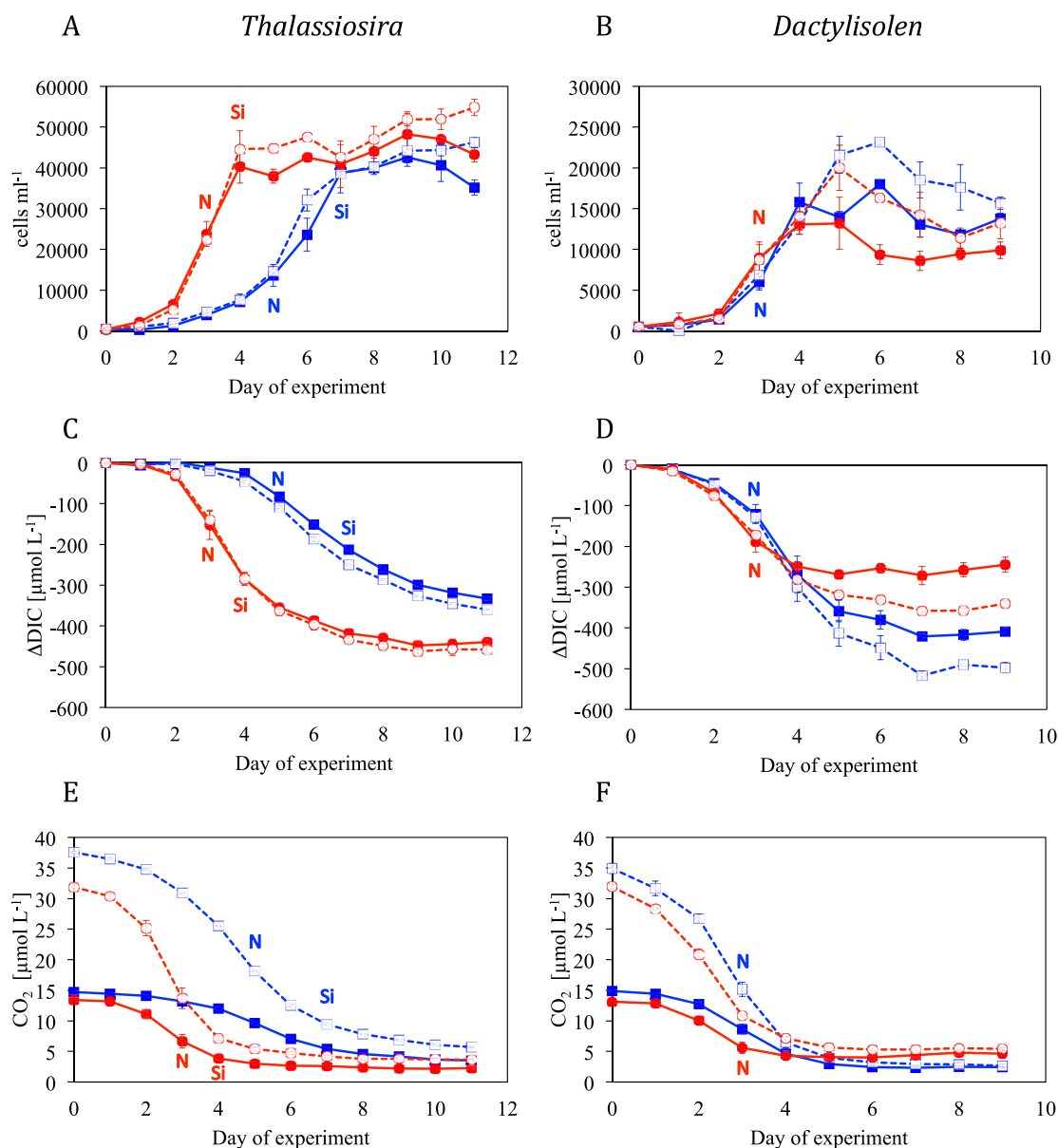
The temperature gradient experiment revealed a significant effect of temperature on growth rate of both diatom species (Fig. 1A). *T. weissflogii* displayed a positive response over the entire range of temperatures from 10°C to 22.5°C, with an optimum at  $\sim 20^\circ\text{C}$ . In contrast, growth rate and TOC build-up by *D. fragilissimus* increased over the range from 10°C to 15°C but decreased beyond that, suggesting an optimum temperature of  $\sim 15^\circ\text{C}$ . Net build-up of organic carbon (TOC = POC + DOC) including the stationary phase (until four days after N-depletion) displayed a similar response to temperature (Fig. 1B).

#### Main experiments

The development of the diatom cultures in both experiments was characterized by a rapid decline in dissolved inorganic nutrients, drawdown of DIC, decrease in  $p\text{CO}_2$  and dissolved CO<sub>2</sub>, and build-up of particulate (POM) and DOM (Figs. 2-4).

#### Growth and nutrient uptake

Initial concentrations of NO<sub>3</sub><sup>-</sup> were 15 μmol L<sup>-1</sup> in both experiments. In the experiment with *T. weissflogii*, NO<sub>3</sub><sup>-</sup> was depleted faster at higher temperatures, reaching concentrations close to zero at t3 (20°C) and t5 (15°C) in both  $p\text{CO}_2$  treatments (indicated in Figs. 2, 3). In the experiment with *D. fragilissimus*, NO<sub>3</sub> depletion occurred on t3 in all temperature and  $p\text{CO}_2$  treatments. PO<sub>4</sub><sup>3-</sup> dropped from initial concentrations of 6 μmol L<sup>-1</sup> and 9 μmol L<sup>-1</sup> in the experiments with *T. weissflogii* and *D. fragilissimus*, respectively, but always remained  $> 2 \mu\text{mol L}^{-1}$  and thus well above

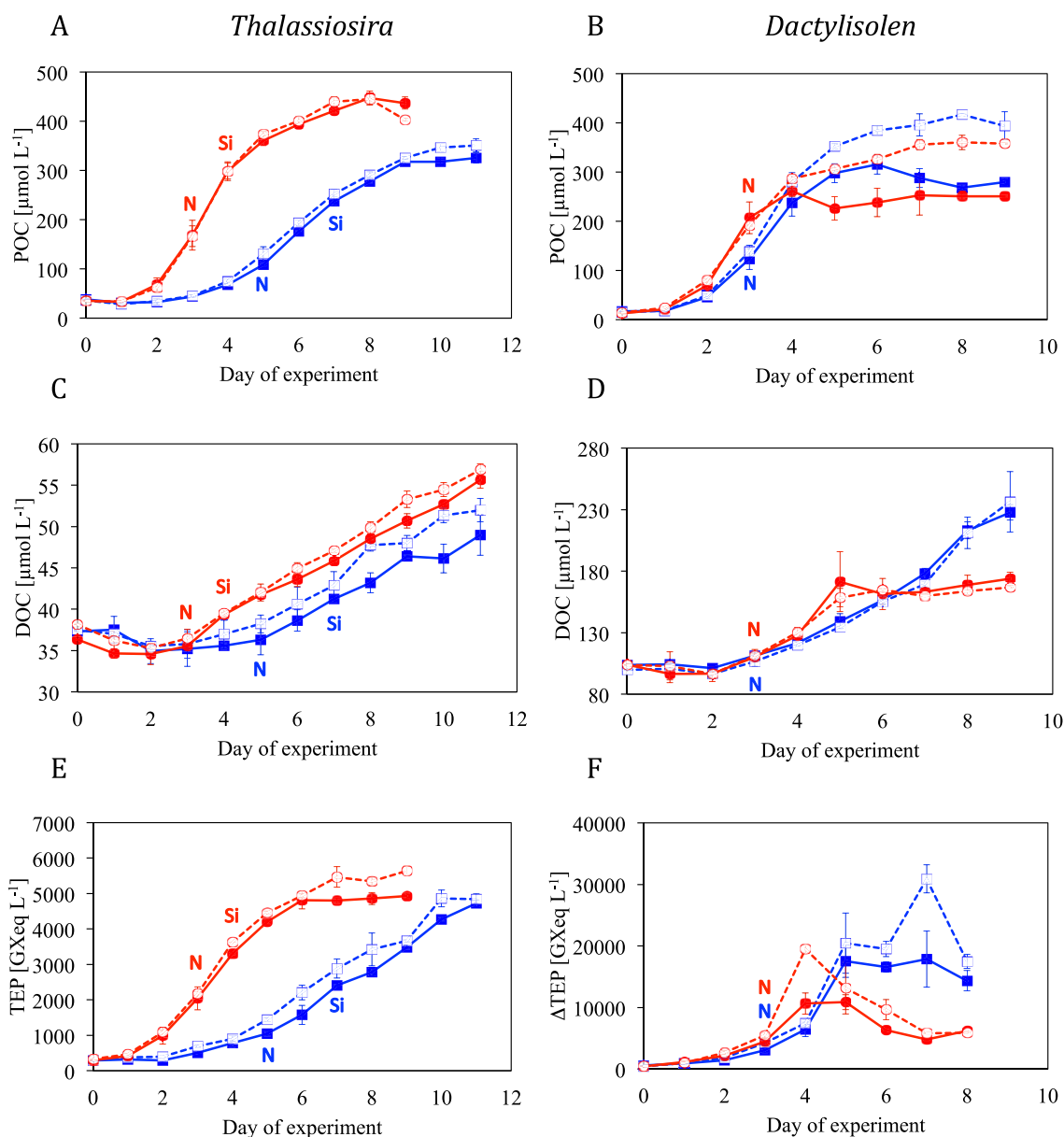


**Fig. 2.** Cell numbers and carbonate system variables in the experiment with *T. weissflogii* (left) and *D. fragilissimus* (right). Shown are temporal development of (A, B) cell concentrations, (C, D) DIC uptake relative to initial values ( $\mu\text{mol L}^{-1}$ ) and (E, F) concentrations of dissolved CO<sub>2</sub> ( $\mu\text{mol L}^{-1}$ ) at 15°C (blue squares) and 20°C (red circles), and in the ambient CO<sub>2</sub> (solid line, closed symbols) and high CO<sub>2</sub> treatments (dashed line, open symbols). Vertical error bars denote range of replicates within each treatment ( $n = 2$ ). Colored letters denote day when cultures reached depletion of inorganic nitrogen (N) and silica (Si) in the 15°C (blue) and 20°C (red) treatments.

depletion. While Si remained replete in the *D. fragilissimus* experiment ( $\sim 40 \mu\text{mol Si(OH)}_4 \text{ L}^{-1}$ ), it reached very low concentrations ( $< 2 \mu\text{mol Si(OH)}_4 \text{ L}^{-1}$ ) in the *T. weissflogii* experiment on t4 and t7 at 20°C and 15°C in both  $p\text{CO}_2$  treatments, respectively. No effect of  $p\text{CO}_2$  on the rate and magnitude of nutrient uptake could be detected in either of the experiments.

The drawdown of nutrients was reflected in rapid growth and an increase in cell concentrations in both experiments (Fig. 2A,B). In the *T. weissflogii* experiment, cell numbers

increased from initial values of  $\sim 500$  to average concentrations of  $33,491 \pm 1646$  (ambient),  $39,717 \pm 1985$  (warming),  $37,218 \pm 2112$  (high CO<sub>2</sub>) and  $42,950 \pm 3968$  (combined) cells mL<sup>-1</sup> in the stationary phase. In the experiment with *D. fragilissimus*, cell concentrations reached average values of  $13,009 \pm 40$  (ambient),  $10,149 \pm 387$  (warming),  $16,477 \pm 137$  (high CO<sub>2</sub>), and  $13,991 \pm 1009$  (combined) cells mL<sup>-1</sup> in the stationary phase. Thus, temperature had a significant effect on average cell concentrations in both experiments. High initial CO<sub>2</sub> resulted in elevated cell numbers in both diatom



**Fig. 3.** Carbon partitioning by *T. weissflogii* (left) and *D. fragilissimus* (right). Shown are temporal development of measured (A, B) POC and (C, D) DOC concentrations ( $\mu\text{mol L}^{-1}$ ), and (E, F) TEP concentrations ( $\mu\text{g GXeq L}^{-1}$ ) in the experiment with *T. weissflogii* (left) and *D. fragilissimus* (right). Style and color-coding as in Fig. 2.

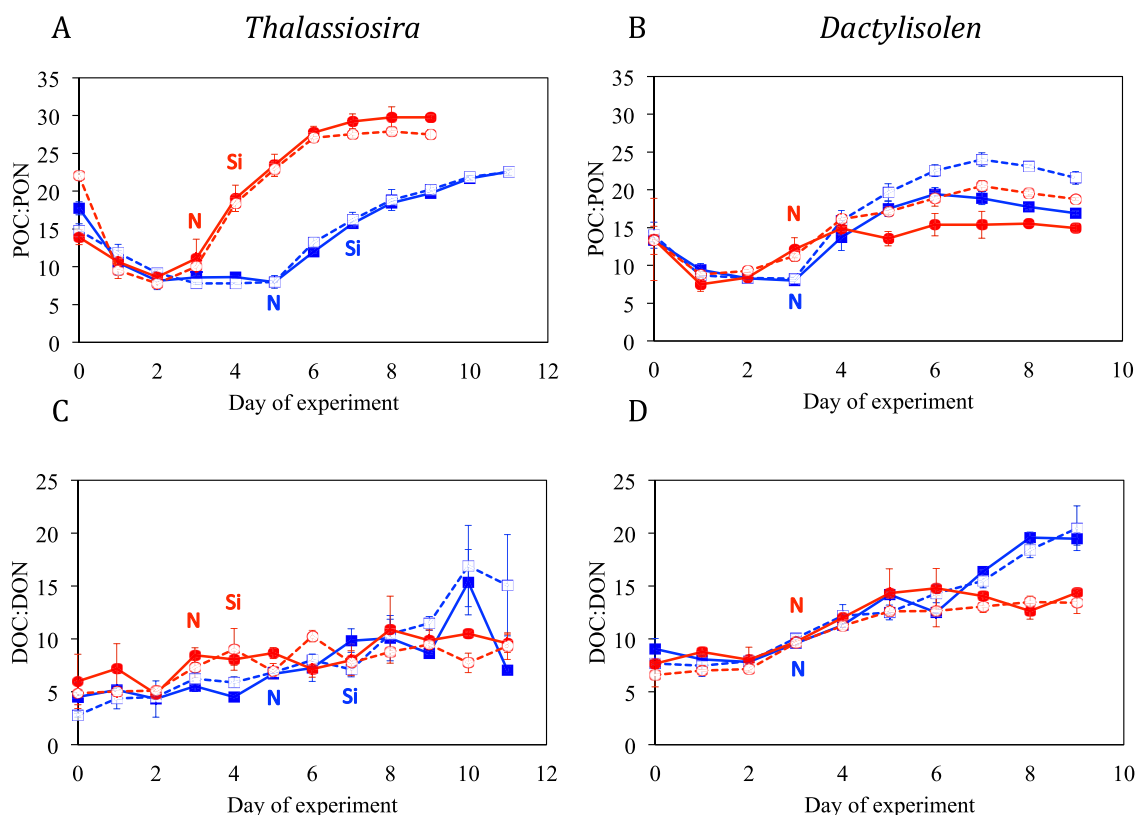
species, however, the effect was only statistically significant for *D. fragilissimus* and not for *T. weissflogii* (Tables 1, 2).

**DIC uptake and build-up of POC, DOC, and TEP**

DIC uptake showed a significant response to temperature and  $p\text{CO}_2$  in both experiments (Fig. 2C,D; Tables 1, 2). Maximum uptake of DIC by *T. weissflogii* amounted to  $333.2 \pm 3.2$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $360.2 \pm 0.8$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $448.1 \pm 9.0$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $462.7 \pm 9.6$  (combined)  $\mu\text{mol C L}^{-1}$ . In the *D. fragilissimus* experiment, maximum drawdown of DIC reached

$420.7 \pm 8.4$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $516.9 \pm 10.7$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $271.6 \pm 11.6$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $358.4 \pm 9.1$  (combined)  $\mu\text{mol C L}^{-1}$ . Elevated  $p\text{CO}_2$  enhanced biologically mediated drawdown of DIC by both species, even though the magnitude of the effect was much larger for *D. fragilissimus* (increase by 22–39%) than for *T. weissflogii* (increase by 3–8%).

The temperature increase from 15°C to 20°C significantly accelerated and enhanced DIC uptake by *T. weissflogii*. This response was more complex in the experiment with *D. fragilissimus*, where elevated temperatures led to a significant



**Fig. 4.** Temporal development of C : N in (A, B) particulate and (C, D) dissolved organic matter in the experiment with *T. weissflogii* (left) and *D. fragilissimus* (right). Style and color-coding as in Fig. 2.

increase ( $p < 0.05$ ) in DIC drawdown under nutrient-replete conditions, but had a negative effect once nitrogen was depleted (Fig. 2B).

The drawdown of DIC was reflected in a concomitant build-up of POC (Fig. 3A,B). In the *T. weissflogii* experiment, maximum POC build-up amounted to  $288.7 \pm 8.3$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $314.7 \pm 13.0$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $412.0 \pm 14.0$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $418.1 \pm 3.5$  (combined)  $\mu\text{mol C L}^{-1}$ . POC build-up was enhanced at higher temperatures, whereas higher initial  $p\text{CO}_2$  levels had no statistically significant effect (Fig. 3A; Table 1). Build-up of POC by *D. fragilissimus* reached maximum values of  $298.3 \pm 19.5$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $402.5 \pm 2.1$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $248.7 \pm 10.9$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $346.0 \pm 4.3$  (combined)  $\mu\text{mol C L}^{-1}$ . Thus, both temperature and  $p\text{CO}_2$  had a significant effect on maximum POC build-up, resulting in a decrease at temperatures beyond its optimum, but an increase at elevated  $p\text{CO}_2$  (Fig. 3B; Table 2). Similar to DIC uptake, temperature had a significant positive effect on POC build-up before N-depletion ( $p < 0.05$ ), but a negative effect afterward.

At the same time, accumulation of DOC was observed over the course of both experiments (Fig. 3C,D). In the *T. weissflogii* experiment, maximum DOC build-up amounted

to  $11.6 \pm 2.4$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $14.5 \pm 1.4$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $19.3 \pm 1.0$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $18.7 \pm 0.7$  (combined)  $\mu\text{mol C L}^{-1}$ . Maximum accumulation of DOC was much greater in the *D. fragilissimus* experiment, reaching  $123.9 \pm 6.2$  (ambient)  $\mu\text{mol C L}^{-1}$ ,  $136.4 \pm 24.6$  (high CO<sub>2</sub>)  $\mu\text{mol C L}^{-1}$ ,  $70.0 \pm 8.3$  (warming)  $\mu\text{mol C L}^{-1}$ , and  $63.2 \pm 0.3$  (combined)  $\mu\text{mol C L}^{-1}$ . Consequently, elevated temperatures had a significant effect on DOC production by both species, resulting in an increase by *T. weissflogii*, but a decrease by *D. fragilissimus*. CO<sub>2</sub> did not have a statistically significant effect on DOC build-up by either of the species (Tables 1, 2).

A notable accumulation of TEP accompanied the accumulation of biomass in both experiments. In the *T. weissflogii* experiment, average TEP accumulation in the stationary phase after N-depletion amounted to  $\sim 2900 \pm 80$  (ambient)  $\mu\text{g GXEq L}^{-1}$ ,  $3330 \pm 180$  (high CO<sub>2</sub>)  $\mu\text{g GXEq L}^{-1}$ ,  $4130 \pm 50$  (warming)  $\mu\text{g GXEq L}^{-1}$ , and  $4520 \pm 50$  (combined)  $\mu\text{g GXEq L}^{-1}$ . Thus, increases in temperature and CO<sub>2</sub> both significantly enhanced TEP accumulation (Fig. 3E; Table 1). The production of TEP by *D. fragilissimus* was generally much higher than that of *T. weissflogii*, reaching average concentrations of  $\sim 11,800 \pm 1520$  (ambient)  $\mu\text{g GXEq L}^{-1}$ ,  $15,660 \pm 1060$  (high CO<sub>2</sub>)  $\mu\text{g GXEq L}^{-1}$ ,  $7940 \pm 510$



**Table 1.** Effects of temperature and CO<sub>2</sub> on biogeochemical parameters in the experiment with *T. weissflogii*. Shown are maximum net uptake of DIC, maximum net build-up of POC, and DOC ( $\mu\text{mol C L}^{-1}$ ), maximum ratios of DIC : NO<sub>3</sub> uptake and POC : PON build-up, as well as average cell numbers ( $\text{mL}^{-1}$ ) and average TEP concentrations ( $\mu\text{g GXeq L}^{-1}$ ) in the stationary phase with respective standard error (SE) of replicates ( $n = 2$ ). Effects of temperature and CO<sub>2</sub> were assessed by PERMANOVA. Significant effects are in bold ( $n = 2$ ,  $df = 1$ ).

	Value ( $\pm$ SE)	Effect	SS	Pseudo-F	p (perm)
$\Delta\text{DIC}_{\text{max}}$					
Ambient	333.2 $\pm$ 3.2				
High Temp.	448.1 $\pm$ 9.0	Temp.	14.8202	287.725	<b>0.004</b>
High CO <sub>2</sub>	360.2 $\pm$ 0.8	CO <sub>2</sub>	0.5687	11.041	<b>0.036</b>
Combined	462.7 $\pm$ 9.6	Temp. $\times$ CO <sub>2</sub>	0.0740	1.438	0.360
$\Delta\text{POC}_{\text{max}}$					
Ambient	288.7 $\pm$ 8.3				
High Temp.	412.0 $\pm$ 14.0	Temp.	18.1083	112.407	<b>0.008</b>
High CO <sub>2</sub>	314.7 $\pm$ 13.1	CO <sub>2</sub>	0.4054	2.517	0.179
Combined	418.1 $\pm$ 3.5	Temp. $\times$ CO <sub>2</sub>	0.1755	1.089	0.425
$\Delta\text{DOC}_{\text{max}}$					
Ambient	11.6 $\pm$ 2.4				
High Temp.	19.3 $\pm$ 1.0	Temp.	1.1596	12.605	<b>0.013</b>
High CO <sub>2</sub>	14.5 $\pm$ 1.4	CO <sub>2</sub>	0.0572	0.621	0.485
Combined	18.7 $\pm$ 0.7	Temp. $\times$ CO <sub>2</sub>	0.1140	1.239	0.345
$[\Delta\text{DIC} : \Delta\text{NO}_3]_{\text{max}}$					
Ambient	22.2 $\pm$ 0.2				
High Temp.	29.9 $\pm$ 0.6	Temp.	0.9880	287.631	<b>0.005</b>
High CO <sub>2</sub>	24.0 $\pm$ 0.1	CO <sub>2</sub>	0.0379	11.036	<b>0.034</b>
Combined	30.8 $\pm$ 0.6	Temp. $\times$ CO <sub>2</sub>	0.0049	1.435	0.354
$[\text{POC} : \text{PON}]_{\text{max}}$					
Ambient	22.6 $\pm$ 0.1				
High Temp.	30.6 $\pm$ 0.5	Temp.	0.8699	274.266	<b>0.007</b>
High CO <sub>2</sub>	22.5 $\pm$ 0.4	CO <sub>2</sub>	0.0335	10.559	<b>0.037</b>
Combined	27.9 $\pm$ 0.5	Temp. $\times$ CO <sub>2</sub>	0.0290	9.131	<b>0.039</b>
Cell number <sub>avg-stat</sub>					
Ambient	33491 $\pm$ 1646				
High Temp.	39717 $\pm$ 1985	Temp.	469.3316	10.072	<b>0.042</b>
High CO <sub>2</sub>	37218 $\pm$ 2112	CO <sub>2</sub>	159.5232	3.423	0.131
Combined	42950 $\pm$ 3968	Temp. $\times$ CO <sub>2</sub>	1.8382	0.039	0.818
TEP <sub>avg-stat</sub>					
Ambient	2898.7 $\pm$ 84.2				
High Temp.	4133.6 $\pm$ 49.3	Temp.	199.6363	116.221	<b>0.003</b>
High CO <sub>2</sub>	3333.2 $\pm$ 183.8	CO <sub>2</sub>	23.2679	13.546	<b>0.031</b>
Combined	4520.7 $\pm$ 50.8	Temp. $\times$ CO <sub>2</sub>	0.4369	0.254	0.607

(warming)  $\mu\text{g GXeq L}^{-1}$ , and 10,760  $\pm$  1380 (combined)  $\mu\text{g GXeq L}^{-1}$  in the stationary phase. Thus, average TEP concentrations in the stationary phase were significantly decreased at higher temperatures, but elevated in the high CO<sub>2</sub> treatments (Fig. 3F; Table 2).

### Stoichiometry

While the same amount of NO<sub>3</sub><sup>-</sup> was consumed in all treatments and in both experiments, the uptake of DIC differed significantly across treatments and between experi-

ments. In the *T. weissflogii* experiment, the uptake ratio of DIC over NO<sub>3</sub><sup>-</sup> was significantly elevated in the treatments at high temperature and elevated  $p\text{CO}_2$ . In contrast, the consumption of carbon relative to nitrogen by *D. fragilissimus* decreased at higher temperatures, but was enhanced in the high CO<sub>2</sub> treatments (Tables 1, 2).

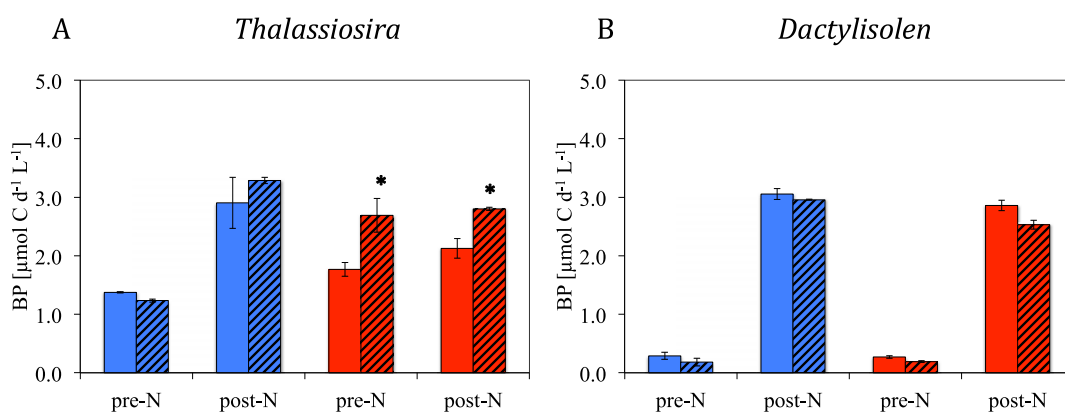
This elevated consumption of carbon over nitrogen is also reflected in the elemental ratios of produced organic matter (Fig. 4). The molar ratio of carbon to nitrogen (C : N) of POM under N-replete conditions remained between 8.0 and

**Table 2.** Effects of temperature and CO<sub>2</sub> on biogeochemical parameters in the experiment with *D. fragilissimus*. Shown are maximum net uptake of DIC, maximum net build-up of POC, and DOC ( $\mu\text{mol C L}^{-1}$ ), maximum ratios of DIC : NO<sub>3</sub> uptake and POC : PON build-up, as well as average cell numbers ( $\text{mL}^{-1}$ ) and average TEP concentrations ( $\mu\text{g GXeq L}^{-1}$ ) in the stationary phase with respective standard error (SE) of replicates ( $n = 2$ ). Effects of temperature and CO<sub>2</sub> were assessed by PERMANOVA. Significant effects are in bold ( $n = 2$ ,  $df = 1$ ).

	Value ( $\pm$ SE)	Effect	SS	Pseudo-F	p(perm)
$\Delta\text{DIC}_{\text{max}}$					
Ambient	420.7 $\pm$ 8.4				
High Temp.	271.6 $\pm$ 11.6	Temp.	28.0617	204.642	<b>0.003</b>
High CO <sub>2</sub>	516.9 $\pm$ 10.7	CO <sub>2</sub>	9.4978	69.264	<b>0.006</b>
Combined	358.4 $\pm$ 9.1	Temp. $\times$ CO <sub>2</sub>	0.0012	0.009	0.869
$\Delta\text{POC}_{\text{max}}$					
Ambient	298.3 $\pm$ 19.5				
High Temp.	248.7 $\pm$ 10.9	Temp.	2.2055	9.895	<b>0.045</b>
High CO <sub>2</sub>	402.5 $\pm$ 2.1	CO <sub>2</sub>	14.1741	63.593	<b>0.005</b>
Combined	346.0 $\pm$ 4.3	Temp. $\times$ CO <sub>2</sub>	0.0958	0.429	0.538
$\Delta\text{DOC}_{\text{max}}$					
Ambient	123.9 $\pm$ 6.2				
High Temp.	70.0 $\pm$ 8.3	Temp.	16.0904	22.938	<b>0.016</b>
High CO <sub>2</sub>	136.4 $\pm$ 24.5	CO <sub>2</sub>	0.2271	0.324	0.595
Combined	63.2 $\pm$ 0.3	Temp. $\times$ CO <sub>2</sub>	1.4160	2.019	0.215
$[\Delta\text{DIC} : \Delta\text{NO}_3]_{\text{max}}$					
Ambient	28.0 $\pm$ 0.6				
High Temp.	18.1 $\pm$ 0.8	Temp.	1.8708	204.680	<b>0.003</b>
High CO <sub>2</sub>	34.5 $\pm$ 0.9	CO <sub>2</sub>	0.6331	69.270	<b>0.006</b>
Combined	23.9 $\pm$ 0.6	Temp. $\times$ CO <sub>2</sub>	8.30e <sup>-5</sup>	0.009	0.875
$[\text{POC} : \text{PON}]_{\text{max}}$					
Ambient	19.4 $\pm$ 0.9				
High Temp.	16.5 $\pm$ 0.7	Temp.	0.2545	17.128	<b>0.018</b>
High CO <sub>2</sub>	24.0 $\pm$ 0.9	CO <sub>2</sub>	0.4724	31.790	<b>0.004</b>
Combined	20.6 $\pm$ 0.6	Temp. $\times$ CO <sub>2</sub>	6.16e <sup>-5</sup>	0.004	0.908
Cell number <sub>avg-stat</sub>					
Ambient	13009 $\pm$ 40				
High Temp.	10149 $\pm$ 387	Temp.	275.8384	24.846	<b>0.016</b>
High CO <sub>2</sub>	16477 $\pm$ 137	CO <sub>2</sub>	505.2507	45.510	<b>0.005</b>
Combined	13991 $\pm$ 1009	Temp. $\times$ CO <sub>2</sub>	5.0633	0.456	0.503
TEP <sub>avg-stat</sub>					
Ambient	11795.8 $\pm$ 1525.3				
High Temp.	7944.4 $\pm$ 513.9	Temp.	833.7093	13.860	<b>0.029</b>
High CO <sub>2</sub>	15658.0 $\pm$ 1062.2	CO <sub>2</sub>	484.1105	8.048	<b>0.049</b>
Combined	10761.0 $\pm$ 1379.1	Temp. $\times$ CO <sub>2</sub>	2.5173	0.042	0.853

9.0 in both experiments, and was thus close to the Redfield value of 6.6. With the depletion of inorganic nitrogen, POC : PON increased rapidly in both experiments. Maximum values of POC : PON in the *T. weissflogii* experiment were significantly elevated at higher temperatures, whereas the effect of CO<sub>2</sub> was small and inconsistent at 15°C (Fig. 4A; Table 1). In contrast, POC : PON values in the *D. fragilissimus* experiment display a decrease at higher temperatures, but an increase at elevated  $p\text{CO}_2$  (Fig. 4B; Table 2).

The C : N ratio of DOM was  $\sim 5.0$  and  $8.0$  in the beginning of the experiment with *T. weissflogii* and *D. fragilissimus*, respectively. After N-depletion, DOC : DON slightly increased in the *T. weissflogii* experiment, however, without a detectable effect of temperature or  $p\text{CO}_2$  (Fig. 4C). In the experiment with *D. fragilissimus*, DOC : DON was significantly elevated at lower temperatures, whereas an effect of  $p\text{CO}_2$  could not be observed at either temperature (Fig. 4D).



**Fig. 5.** Average rates of net BP in the period before and after nitrogen depletion (denoted “pre-N” and “post-N,” respectively) in the experiment with (A) *T. weissflogii* and (B) *D. fragilissimus* at 15 (blue) and 20°C (red), and in the ambient (solid) and high CO<sub>2</sub> treatment (hatched). Vertical error bars denote range of replicates within each treatment ( $n = 2$ ). Asterisks denote significant effect of CO<sub>2</sub> in the respective temperature treatment ( $p < 0.05$ ).

### Bacterial activity

In the experiment with *T. weissflogii*, net BP in the 15°C treatments amounted to  $1.30 \pm 0.4 \mu\text{mol C L}^{-1} \text{d}^{-1}$  under N-replete conditions and increased to  $3.09 \pm 0.21 \mu\text{mol C L}^{-1} \text{d}^{-1}$  under N-depletion, irrespective of CO<sub>2</sub>. In the 20°C treatments, however, BP was elevated under high CO<sub>2</sub> conditions displaying values of  $1.77 \pm 0.12 \mu\text{mol C L}^{-1} \text{d}^{-1}$  (ambient CO<sub>2</sub>) and  $2.69 \pm 0.29 \mu\text{mol C L}^{-1} \text{d}^{-1}$  (high CO<sub>2</sub>) during N-repletion and  $2.13 \pm 0.17 \mu\text{mol C L}^{-1} \text{d}^{-1}$  (ambient CO<sub>2</sub>) and  $2.80 \pm 0.03 \mu\text{mol C L}^{-1} \text{d}^{-1}$  (high CO<sub>2</sub>) in the period after N-depletion. A significant effect of temperature or CO<sub>2</sub> could not be detected when PERMANOVA was applied to both temperatures. However, CO<sub>2</sub> significantly enhanced BP in the *T. weissflogii* cultures at 20°C ( $p < 0.05$ ) (Fig. 5A).

In the experiment with *D. fragilissimus*, BP increased from initial levels of  $0.23 \pm 0.02 \mu\text{mol C L}^{-1} \text{d}^{-1}$  under N-replete conditions to  $2.85 \pm 0.08 \mu\text{mol C L}^{-1} \text{d}^{-1}$  in the period after N-depletion, showing no significant difference between temperature and CO<sub>2</sub> treatments (Fig. 5B).

### Discussion

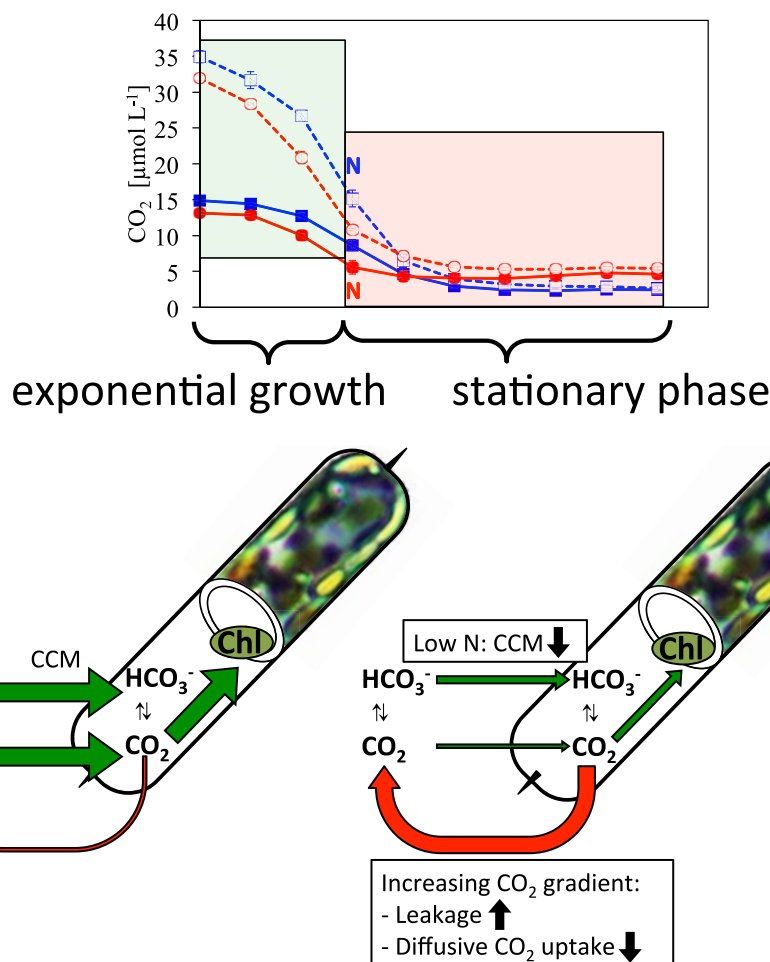
Our results demonstrate that carbon uptake of the diatoms *T. weissflogii* and *D. fragilissimus* is markedly sensitive to CO<sub>2</sub> and temperature, and that this sensitivity is mainly driven by excess carbon fixation in the stationary phase under N-depletion. Carbon overconsumption has been observed in previous experiments and field studies (Banse 1994; Körtzinger et al. 2001; Sambrotto et al. 1993) and is usually associated with nutrient stress (Biddanda and Benner 1997; Wetz and Wheeler 2003). This is consistent with our experiments, where a substantial portion of inorganic carbon uptake and build-up of organic matter (up to 70%) by the diatom cultures occurred in the stationary phase: DIC uptake continued for several days after N-depletion and was even higher than during the period when inor-

ganic N was available (in absolute terms), suggesting a strong decoupling of carbon and nitrogen cycling (Fig. 2C,D). Even though diatoms are known for luxury consumption of nutrients, i.e., storage inside their vacuole and delayed cell division and biomass production (Goldman et al. 1979), the magnitude of the stoichiometric decoupling observed in our experiments is remarkable. Interestingly, the sensitivity to CO<sub>2</sub> and temperature was most prominent in this period of N-limitation. Possible mechanisms behind that will be discussed below.

Note that *T. weissflogii* probably experienced colimitation by N and Si later in the experiment, whereas *D. fragilissimus* remained only N-limited throughout the experiment (Figs. 1-3). As Si is essential for cell division, this might have affected the biomass yield of *T. weissflogii* in the stationary phase and obscured possible effects of temperature and CO<sub>2</sub> on carbon uptake and partitioning in this experiment.

### CO<sub>2</sub> effect on carbon uptake

Cumulative uptake of DIC by both diatom species was enhanced in the high CO<sub>2</sub> treatments, with the effect being much stronger on *D. fragilissimus* (+39%) than on *T. weissflogii* (+8%). This observation is in line with an earlier mesocosm experiment where DIC drawdown of a natural diatom-dominated community showed a marked increase under elevated  $p\text{CO}_2$  (Riebesell et al. 2007). Remarkably, the fertilizing effect of CO<sub>2</sub> in our experiments was not detectable before N-depletion and when  $p\text{CO}_2$  was still close to target levels of 400  $\mu\text{atm}$  and 1000  $\mu\text{atm}$ . Instead, most of the differences in DIC uptake occurred after N-depletion and when dissolved CO<sub>2</sub> dropped below  $\sim 10 \mu\text{mol L}^{-1}$ , suggesting that inhibition of carbon assimilation at low concentrations of dissolved CO<sub>2</sub> played an important role in the observed response (Fig. 2C-F). This is in line with previous studies, which demonstrated that carbon acquisition in diatoms is highly sensitive to CO<sub>2</sub>, especially at low concentrations of



**Fig. 6.** Schematic diagram illustrating the influence of seawater CO<sub>2</sub> on carbon uptake by the diatoms over the course of the experiments. In the beginning of the experiment, CO<sub>2</sub> concentrations are relatively high and nutrients are available, leading to exponential growth of the diatoms (green square). Inorganic carbon is taken up either passively via diffusion of CO<sub>2</sub> or actively through a CCM, which transports HCO<sub>3</sub><sup>-</sup> into the cell where it is subsequently converted to CO<sub>2</sub> and used for photosynthesis inside the chloroplast. Over the course of the experiment, CO<sub>2</sub> concentrations become very low and nitrogen gets depleted. In this stationary phase with low CO<sub>2</sub> and N-depletion, the CO<sub>2</sub> gradient between the cell interior and the surrounding seawater becomes so large that diffusive CO<sub>2</sub> uptake ceases and diffusive leakage of CO<sub>2</sub> becomes dominant. At the same time, active carbon uptake through the CCM decreases sharply, as there is a lack in nitrogen for the synthesis of CCM-specific proteins. Altogether, this leads to a situation in which the cells are no longer able to achieve a net uptake of inorganic carbon. Higher initial CO<sub>2</sub> conditions allowed the diatoms to take up more DIC until they reached such low CO<sub>2</sub> concentrations where carbon limitation occurred.

dissolved CO<sub>2</sub> (Riebesell et al. 1993; Burkhardt et al. 1999; Rost et al. 2003). Other studies also reported a close correlation of growth rate and POC production with dissolved CO<sub>2</sub>, suggesting growth inhibition below 5–7.5 μmol CO<sub>2</sub> (Engel 2002; Bach et al. 2013).

**Possible mechanisms behind the CO<sub>2</sub> effect**

In our experiments, two mechanisms might have contributed to inhibition of carbon uptake at low CO<sub>2</sub>: diffusive leakage of CO<sub>2</sub> from the cell and N-deficiency affecting the performance of the CCM.

Most phytoplankton species can actively take up HCO<sub>3</sub><sup>-</sup> in addition to passive uptake of CO<sub>2</sub> via diffusion through operation of a CCM. Thereby, they enhance their intracellu-

lar CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup>, which creates a concentration gradient between inside and outside the cell. However, photosynthetic carbon fixation by RubisCO is restricted to CO<sub>2</sub>, which is why accumulated HCO<sub>3</sub><sup>-</sup> has to be converted to CO<sub>2</sub> in the cell interior (Rost et al. 2003; Hopkinson et al. 2011). Thus, at decreasing concentrations of dissolved CO<sub>2</sub>, the diffusion gradient between the cell interior and surrounding seawater increases strongly and CO<sub>2</sub> leakage becomes more and more dominant (Rost et al. 2006; Hopkinson et al. 2011; Bach et al. 2013).

At the same time, inhibition of carbon uptake at low CO<sub>2</sub> was likely also linked to N-limitation. As diffusive uptake of carbon becomes less efficient at lower CO<sub>2</sub> in seawater,

active carbon acquisition through a CCM gains importance. Diatoms possess a highly regulated and efficient CCM compared with some other phytoplankton groups (Burkhardt et al. 2001; Rost et al. 2003; Reinfelder 2011). However, the functioning of the CCM is ultimately controlled by the availability of nitrogen, which is required for the synthesis of specific proteins used in the CCM and can therefore be severely compromised by N-limitation (Geider et al. 1993; Giordano et al. 2005). With respect to our experiments, this implies that carbon limitation of the diatoms in the stationary phase was intensified by N-deficiency. As diatoms rely heavily on an efficient CCM for carbon acquisition, this effect became particularly pronounced at low CO<sub>2</sub> concentrations in seawater.

Altogether, these considerations indicate that diatoms in the high pCO<sub>2</sub> treatments could take up more DIC until they reached such low CO<sub>2</sub> concentrations, where carbon limitation occurred through the balance between uptake and leakage of inorganic carbon, and the exacerbating effect of N-limitation on the CCM (Fig. 6).

Interestingly, cell abundances were elevated in the high CO<sub>2</sub> treatments, with the effect being much more pronounced on *D. fragilissimus* (+40% cell concentrations) than on *T. weissflogii* (+10%) (Fig. 2A,B). This indicates that the observed CO<sub>2</sub> effect on DIC uptake stems from an increase in the carrying capacity of the system under high CO<sub>2</sub> rather than from a physiological response at the cellular level. Indeed, our data suggest that DIC uptake on a per cell basis was not considerably affected by CO<sub>2</sub> (calculated data not shown, see Fig. 2). Possibly, high CO<sub>2</sub> conditions resulted in a “memory effect” on the diatom cultures: higher CO<sub>2</sub> concentrations could have facilitated carbon acquisition to an extent that cells invested less resources in a CCM during the first few days of the experiment, and in turn had more resources left (for growth and cell division) in the phase of N-depletion and low CO<sub>2</sub> when carbon acquisition was complicated by the mechanism explained above. Thus, such a “memory effect” could explain the observation that although the differences in the carbonate systems of both treatments were largest during the first few days of the experiments, treatment effects became prominent much later when the differences between CO<sub>2</sub> in both treatments were small.

#### Differences among the species

The response to high CO<sub>2</sub> differed greatly between *T. weissflogii* and *D. fragilissimus*. While it cannot be excluded that this is partly due to secondary Si-limitation of *T. weissflogii*, it is likely that these CO<sub>2</sub>-associated differences are either species-specific or related to cell size in general.

Species-specific differences seem obvious as photosynthetic pathways of diatoms are diverse and there are substantial differences regarding the preference for the form of inorganic carbon (CO<sub>2</sub> or HCO<sub>3</sub><sup>-</sup>) among taxa (Rost et al. 2003; Giordano et al. 2005; Roberts et al. 2007). Considering that under ocean acidification conditions changes in dis-

solved CO<sub>2</sub> are much larger than those in HCO<sub>3</sub><sup>-</sup>, it is likely that species more dependent on CO<sub>2</sub> as a carbon source, are more affected by changing pCO<sub>2</sub>. Such species-specific variations could explain why *D. fragilissimus* displayed a much stronger reaction to high pCO<sub>2</sub> than *T. weissflogii*. The fact that the latter species did not show a strong sensitivity to CO<sub>2</sub> might also be linked to a C<sub>4</sub> pathway in their CCM. Previous work has demonstrated that *T. weissflogii* (and possibly other species) apply a biochemical CCM where HCO<sub>3</sub><sup>-</sup> is transported into the cell and converted into an organic C<sub>4</sub> compound by the C<sub>4</sub>-carboxylase PEPCase prior to fixation by RubisCO (Reinfelder et al. 2000; Roberts et al. 2007).

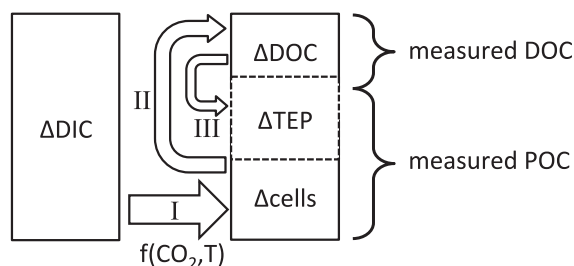
Besides species-specific differences, the diverging response of the two species examined here could be related to cell size of the diatoms. In our study, the effect of CO<sub>2</sub> was much more prominent for the large, chain-forming *D. fragilissimus*, than for the rather small, unicellular *T. weissflogii*. Possibly, *D. fragilissimus* experienced a much stronger carbon limitation at low concentrations of dissolved CO<sub>2</sub> (as explained above) than *T. weissflogii*. Based on theoretical considerations, efficiency of carbon uptake should be related to cell size: small cells have a larger surface area per unit volume than large cells and can thus support more transporters on the cell surface relative to their volume and maintain a thinner diffusion boundary layer (Wolf-Gladrow and Riebesell 1997; Finkel et al. 2010). In addition, larger cells with lower surface-to-volume ratios have to rely more heavily on a CCM than on diffusive uptake of CO<sub>2</sub> and could consequently be affected more severely by N-limitation of their CCM. These considerations are supported by recent findings from laboratory experiments, which revealed a pronounced size dependence of the growth-stimulating effect of CO<sub>2</sub> on diatoms (Wu et al. 2014). Similar observations have been made on natural, mixed phytoplankton assemblages, e.g., in the Ross Sea, where elevated CO<sub>2</sub> favored large chain-forming diatoms over small single-celled diatoms, resulting in an overall increase in carbon fixation of the phytoplankton community (Tortell et al. 2008).

As mentioned earlier, *T. weissflogii* reached Si-limitation in the stationary phase (Figs. 2, 3). This shortage in Si supply might have prevented cells from continued division, thereby possibly weakening bulk carbon uptake of the cultures and obscuring effects of CO<sub>2</sub> and temperature to a certain degree.

#### Temperature effect on carbon uptake

Temperature had an appreciable effect on the magnitude of DIC uptake and build-up of biomass by both diatom species (Figs. 2, 3). Warming from 15°C to 20°C resulted in a 31% increase in carbon uptake by *T. weissflogii*, but a 33% decrease by *D. fragilissimus*. This response is consistent with the results from the temperature-gradient experiments, which suggest different physiological optimum temperatures for the two species (Fig. 1). Consequently, the temperature rise from 15°C to 20°C in the main experiment had a





**Fig. 7.** Schematic diagram showing the influence of partitioning of DIC uptake into the different organic carbon pools. Diatom cultures fix inorganic carbon, which is modulated by CO<sub>2</sub> and temperature (I). With decreasing nutrient availability, an increasing portion of the fixed carbon is exuded as DOC (II). A certain portion of this DOC constitutes precursor material for TEP, which form abiotically (III) and can constitute a major fraction of POC. Measured standing stocks of DOC and POC, however, do not allow for a quantitative discrimination between cellular carbon and TEP or a determination of “true” DOC release that includes dissolved TEP precursor material. Thus, while effects of CO<sub>2</sub> and temperature on net carbon uptake can be easily detected in the data, possible effects on the relative partitioning between the pools are obscured through the various conversion processes between the pools, as well as bacterial degradation (not shown).

stimulating effect on *T. weissflogii*, but acted as a stressor on *D. fragilissimus* and pushed it beyond its temperature optimum.

It is well-known that temperature exerts a substantial influence on most cellular processes, such as enzymatic reactions. Consequently, growth and metabolic rates are enhanced by increasing temperatures until a species’ optimum is reached. A further increase in temperature beyond this optimum is detrimental, leading to reduced growth rates, cellular damage, and eventually cell death (Raven and Geider 1988; Boyd et al. 2010; Finkel et al. 2010). While the effects of temperature on phytoplankton growth rates have been investigated in a number of previous studies (Eppley 1972; Boyd et al. 2013), implications for biogeochemical cycling have received only little attention. Our results show that the direction and magnitude of the effect, e.g., on net carbon uptake strongly depends on the temperature change relative to the temperature range and optimum of the investigated species. In fact, such species-specific differences in temperature sensitivity were identified as the main driver behind seemingly contradictory effects of warming on carbon cycling by natural plankton communities in earlier mesocosm studies (Wohlers et al. 2009; Taucher et al. 2012). These studies demonstrated that the physiological response of the dominant species can determine the temperature effect on the entire plankton community. Another factor that could have possibly affected carbon uptake to in our experiments is the effect of temperature on the buffer capacity of seawater (Sundquist et al. 1979). At 20°C, the same amount of DIC uptake is accompanied by smaller

changes in *p*CO<sub>2</sub> and pH than at 15°C. Thus, an elevated buffer capacity at higher temperatures might increase the amount of DIC that can be taken up by phytoplankton until CO<sub>2</sub> limitation is reached. However, as the temperature response differed severely between the two diatom species in this study, it is likely that the direct physiological effect of temperature was dominant and overshadowed the possible effects of temperature-driven changes in the buffer capacity.

#### Fate of carbon overconsumption: Partitioning of organic matter and the role of TEP

DIC uptake was accompanied by a substantial build-up of organic carbon in both experiments that largely mirrored the respective effects of temperature and CO<sub>2</sub> on the two diatom species (Fig. 3). Our data indicate that the partitioning of fixed carbon between the particulate and dissolved pool as well as the effects of CO<sub>2</sub> and temperature, are tightly linked to the formation of gel-like TEP.

It is well known that carbon overconsumption after nutrient depletion is mainly released as DOC, thereby acting as a “carbon overflow” mechanism that helps to preserve the metabolic functionality of the cell when there is an imbalance between light and nutrients (Fogg 1983). Previous studies have shown that abiotic formation can transform a substantial fraction of DOC exudates into TEP on very short timescales (Passow 2000; Engel et al. 2004). With respect to our experiments, this suggests that a large portion of organic matter that was originally released as DOC comprised TEP-precursors, which were then rapidly transformed into TEP (Fig. 3E,F). This is consistent with previous studies, which demonstrated that TEP can constitute a substantial fraction of total POC (Engel et al. 2004; Wetz and Wheeler 2007). It also illustrates how TEP formation affects the interpretation of organic matter partitioning by underestimating true DOC production and overestimating the amount of organic matter retained as cellular carbon. Due to these mechanisms it cannot be resolved whether the effects of temperature and CO<sub>2</sub> on DIC uptake simply translated into the build-up of the different organic carbon pools, or if there were additional direct effects on the relative partitioning between cellular carbon, DOC and TEP (Fig. 7).

Organic carbon partitioning between the particulate and dissolved phase differed notably among species in the experiments presented here. While inorganic carbon uptake by *T. weissflogii* could almost entirely be traced to the POC pool (~ 95%), a much larger fraction of DIC uptake accumulated as DOC (20–45%) in the experiment with *D. fragilissimus* (Fig. 3). These differences might be attributable to variations in the amount and quality of produced DOC, the fraction of TEP-precursors contained in this DOC, and their subsequent conversion to TEP (Fig. 7). Such differences might also explain why some studies found carbon overconsumption to be channeled mainly into an increase in DOC (Sambrotto et al. 1993; Koeve, 2004), whereas others could trace this

additional carbon almost entirely to POC and TEP (Engel et al. 2002; Wetz and Wheeler 2007).

The substantial formation of TEP, which are very carbon-rich and usually have C : N ratios of >20 (Mari et al. 2001), can explain most of the high POC : PON ratios of up to 25 found in our experiments (Fig. 4). However, changes in the cellular composition might have additionally contributed to the observed increase in POC : PON. Particularly in the experiment with *D. fragilissimus*, maximum cell concentrations varied strongly with CO<sub>2</sub> and temperature, even though the initial amount of inorganic nitrogen was identical. These differences in cell numbers must be linked with changes in their elemental composition, which can be highly variable in diatoms especially under nutrient starvation (Brzezinski 1985; Goldman et al. 1992; Harrison et al. 1977). Such variations can be driven by production of cells with decreasing nitrogen content under N-limitation or an increase in the intracellular content of carbon-enriched compounds, such as lipids and carbohydrates (Goldman et al. 1992; McGinnis et al. 1997; Bach et al. 2013).

Furthermore, it has to be noted that the temporal development of measured DIC, POC, and DOC must be considered as net change. It is likely that actual DOC release was underestimated due to bacterial respiration. In our experiments, we resolved net BP. Even though measured BP did not exceed 3  $\mu\text{mol L}^{-1} \text{d}^{-1}$ , bacterial carbon demand (i.e., the gross flux of carbon through the bacteria pool) could have been higher, depending on the bacterial growth efficiency (BGE). Because DOC build-up by *T. weissflogii* was relatively low (< 20  $\mu\text{mol L}^{-1}$ , Table 1), slight differences in bacterial respiration might have a notable effect on measured DOC accumulation. BP was elevated in the high CO<sub>2</sub> treatments of *T. weissflogii* (Fig. 5A), which is in line with previous studies that found a positive effect of elevated CO<sub>2</sub> on bacterial activity (Grossart et al. 2006; Endres et al. 2014). While we do not have information about BGE, elevated bacterial carbon demand could have obscured the visible effect of high CO<sub>2</sub> on accumulation of DOC in this experiment. In contrast, DOC build-up by *D. fragilissimus* was much higher (~ 60-130  $\mu\text{mol L}^{-1}$ , Table 2) and net BP was similar across all treatments (Fig. 5B). Thus, while heterotrophic bacterial consumption could have led to an underestimate of true DOC production, it is not clear, without more information about BGE, if bacterial carbon demand obfuscated a CO<sub>2</sub> effect on DOC exudation in the *D. fragilissimus* experiment. It should also be noted that observed differences in bacterial activity among treatments might be largely driven by indirect effects, such as differences in the quality of produced substrate by the diatoms (e.g., DOC), than by direct effects of temperature and CO<sub>2</sub> on metabolic rates of bacteria.

Altogether, both the formation of TEP from dissolved precursor material, as well as uncertainties about bacterial carbon demand could have led to an underestimation of true

DOC release and obscured the effects of temperature and CO<sub>2</sub> on carbon partitioning in our experiments (Fig. 7).

### Potential implications for the marine carbon cycle

Our experiments revealed a notable sensitivity of carbon uptake and production of organic matter of two diatom species to both temperature and CO<sub>2</sub>, particularly in the stationary phase after nitrogen was depleted. Remarkably, up to 70% of DIC uptake occurred after N-depletion. Although previous studies concluded that CO<sub>2</sub> is usually not a limiting factor for photosynthetic carbon fixation, with only negligible impacts of elevated CO<sub>2</sub> on diatoms, our results demonstrate that the effect becomes much more prominent when considering temporal bloom dynamics and especially the stationary phase. We show that a higher initial  $p\text{CO}_2$  can substantially increase the amount of inorganic carbon that is taken up over the course of a diatom bloom before CO<sub>2</sub> becomes a limiting factor: net DIC uptake in our experiments was enhanced by 8–39% under high CO<sub>2</sub> conditions. At the same time, our study illustrates that the same magnitude of warming can have either stimulating or inhibiting effects on carbon uptake and production of biomass by diatoms that co-occur in nature, depending on their species-specific temperature range.

The magnitude of these observed responses is quite significant, considering that changes in CO<sub>2</sub> and temperature as in our experiment can be expected by the end of this century. Thus, the observed effects on the diatoms could act as a feedback mechanism in the marine carbon cycle. Whether the sensitivity of carbon uptake to temperature and CO<sub>2</sub> presented in this study results in a negative or positive climate feedback would depend on the fate of the organic carbon. It has been shown that DOC can significantly contribute to the carbon flux to the deep ocean through transport via deep water formation and mixing (Hansell and Carlson 1998; Carlson et al. 2010). An increase in DOC release by phytoplankton could therefore enhance the transport of carbon from the atmosphere to the ocean interior, depending on the quality and bioavailability of the produced DOC (Carlson and Hansell 2014). If the additional DOC gets rapidly remineralized in the surface ocean, the net effect on carbon flux would be zero. Thus, more information on degradability of DOC release by phytoplankton is required. Even stronger effects on the biological pump could be mediated by changes in the production of TEP, which facilitate the formation of marine aggregates through their high stickiness and thus play an important role in particle flux to the ocean interior (Alldredge et al. 1993; Logan et al. 1995; Passow 2002). Our results suggest a substantial sensitivity of TEP formation to temperature and CO<sub>2</sub>: TEP accumulation in the stationary phase was profoundly enhanced under high CO<sub>2</sub> conditions (+12% for *T. weissflogii* and +35% for *D. fragilissimus*) and also displayed a substantial response to warming from 15°C to 20°C (+40% *T. weissflogii* and –30% by *D. fragilissimus*). Such a significant increase in

TEP production could promote the formation of fast-sinking aggregates, thereby enhancing the particle flux out of the euphotic zone, and ultimately the biological pump (Arrigo 2007).

However, sensitivities to temperature and CO<sub>2</sub> seem to be highly variable among species, making it difficult to predict whether the net effect of ocean warming and acidification will be an enhancement of the biological pump or an acceleration of the microbial loop. The outcome for entire pelagic ecosystems will depend on the sensitivities of the different ecosystem components and its most important species, as these effects will ultimately propagate into community composition, food web dynamics, and the efficiency of the biological pump. As the major fraction of DIC uptake and organic matter build-up in our experiments occurred after N-depletion, we suggest that more attention should be paid to the stationary phase and post-bloom dynamics in future studies, especially on the ecosystem level (e.g., mesocosm studies).

## References

- Allredge, A. L., U. Passow, and B. E. Logan. 1993. The abundance and significance of a class of large transparent organic particles in the ocean. *Deep Sea Res. Part I Oceanogr Res. Pap.* **40**: 1131–1140. doi:10.1016/0967-0637(93)90129-q
- Anderson, M. J. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecol.* **26**: 32–46. doi:10.1111/j.1442-9993.2001.01070.pp.x
- Arrigo, K. R. 2007. Carbon cycle - Marine manipulations. *Nature* **450**: 491–492. doi:10.1038/450491a
- Bach, L. T., L. C. M. Mackinder, K. G. Schulz, G. Wheeler, D. C. Schroeder, C. Brownlee, and U. Riebesell. 2013. Dissecting the impact of CO<sub>2</sub> and pH on the mechanisms of photosynthesis and calcification in the coccolithophore *Emiliania huxleyi*. *New Phytol.* **199**: 121–134. doi:10.1111/nph.12225
- Bandstra, L., B. Hales, and T. Takahashi. 2006. High-frequency measurements of total CO<sub>2</sub>: Method development and first oceanographic observations. *Mar. Chem.* **100**: 24–38. doi:10.1016/j.marchem.2005.10.009
- Banse, K. 1994. Uptake of inorganic carbon and nitrate by marine plankton and the Redfield ratio. *Glob. Biogeochem. Cycle* **8**: 81–84. doi:10.1029/93GB02865
- Beardall, J., S. Stojkovic, and S. Larsen. 2009. Living in a high CO<sub>2</sub> world: Impacts of global climate change on marine phytoplankton. *Plant Ecol. Divers.* **2**: 191–205. doi:10.1080/17550870903271363
- Biddanda, B., and R. Benner. 1997. Carbon, nitrogen, and carbohydrate fluxes during the production of particulate and dissolved organic matter by marine phytoplankton. *Limnol. Oceanogr.* **42**: 506–518. doi:10.4319/lo.1997.42.3.0506
- Boyd, P. W., R. Strzepek, F. X. Fu, and D. A. Hutchins. 2010. Environmental control of open-ocean phytoplankton groups: Now and in the future. *Limnol. Oceanogr.* **55**: 1353–1376. doi:10.4319/lo.2010.55.3.1353
- Boyd, P. W., and others. 2013. Marine phytoplankton temperature versus growth responses from polar to tropical waters - Outcome of a scientific community-wide study. *Plos One* **8**: e63091. doi:10.1371/journal.pone.0063091
- Brzezinski, M. A. 1985. The Si-C-N ratio of marine diatoms - Interspecific variability and the effect of some environmental variables. *J. Phycol.* **21**: 347–357. doi:10.1111/j.0022-3646.1985.00347.x
- Buesseler, K. O. 1998. The decoupling of production and particulate export in the surface ocean. *Glob. Biogeochem. Cycle* **12**: 297–310. doi:10.1029/97GB03366
- Burkhardt, S., G. Amoroso, U. Riebesell, and D. Sultemeyer. 2001. CO<sub>2</sub> and HCO<sub>3</sub>-uptake in marine diatoms acclimated to different CO<sub>2</sub> concentrations. *Limnol. Oceanogr.* **46**: 1378–1391. doi:10.4319/lo.2001.46.6.1378
- Burkhardt, S., I. Zondervan, and U. Riebesell. 1999. Effect of CO<sub>2</sub> concentration on C : N : P ratio in marine phytoplankton: A species comparison. *Limnol. Oceanogr.* **44**: 683–690. doi:10.4319/lo.1999.44.3.0683
- Caldeira, K., and M. E. Wickett. 2003. Anthropogenic carbon and ocean pH. *Nature* **425**: 365–365. doi:10.1038/425365a
- Carlson, C. A., and D. A. Hansell. 2014. DOM sources, sinks, reactivity and budgets in the ocean, In D. A. Hansell and C. A. Carlson [eds.], *Biogeochemistry of marine dissolved organic matter* 2nd ed. Academic Press.
- Carlson, C. A., and others. 2010. Dissolved organic carbon export and subsequent remineralization in the mesopelagic and bathypelagic realms of the North Atlantic basin. *Deep Sea Res. Part II Top. Stud. Oceanogr.* **57**: 1433–1445. doi:10.1016/j.dsr2.2010.02.013
- Dickson, A. G., and F. J. Millero. 1987. A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media. *Deep Sea Res. Part A Oceanogr. Res. Pap.* **34**: 1733–1743. doi:10.1016/0198-0149(87)90021-5
- Dickson, A. G., C. L. Sabine, and J. R. Christian. 2007. Guide to best practices for ocean CO<sub>2</sub> measurements - PICES Special Publication 3.
- Doney, S. C., V. J. Fabry, R. A. Feely, and J. A. Kleypas. 2009. Ocean acidification: The other CO<sub>2</sub> problem. *Annu. Rev. Mar. Sci.* **1**: 169–192. doi:10.1146/annurev.marine.010908.163834
- Duarte, C. M. 2007. Marine ecology warms up to theory. *Trends Ecol. Evol.* **22**: 331–333. doi:10.1016/j.tree.2007.04.001
- Egleston, E. S., C. L. Sabine, and F. M. M. Morel. 2010. Revelle revisited: Buffer factors that quantify the response of ocean chemistry to changes in DIC and alkalinity. *Glob.*



- Biogeochem. Cycle **24**: GB1002. doi:10.1029/2008gb003407
- Endres, S., L. Galgani, U. Riebesell, K. G. Schulz, and A. Engel. 2014. Stimulated bacterial growth under elevated pCO<sub>2</sub>: Results from an off-shore mesocosm study. Plos One **9**: e99228. doi:10.1371/journal.pone.0099228
- Engel, A. 2002. Direct relationship between CO<sub>2</sub> uptake and transparent exopolymer particles production in natural phytoplankton. J. Plankton Res. **24**: 49–53. doi:10.1093/plankt/24.1.49
- Engel, A., B. Delille, S. Jacquet, U. Riebesell, E. Rochelle-Newall, A. Terbruggen, and I. Zondervan. 2004. Transparent exopolymer particles and dissolved organic carbon production by *Emiliania huxleyi* exposed to different CO<sub>2</sub> concentrations: A mesocosm experiment. Aquat. Microb. Ecol. **34**: 93–104. doi:10.3354/ame034093
- Engel, A., S. Goldthwait, U. Passow, and A. Alldredge. 2002. Temporal decoupling of carbon and nitrogen dynamics in a mesocosm diatom bloom. Limnol. Oceanogr. **47**: 753–761. doi:10.4319/lo.2002.47.3.0753
- Engel, A., and U. Passow. 2001. Carbon and nitrogen content of transparent exopolymer particles (TEP) in relation to their Alcian Blue adsorption. Mar. Ecol. Prog. Ser. **219**: 1–10. doi:10.3354/meps219001
- Eppley, R. W. 1972. Temperature and phytoplankton growth in sea. Fish. Bull. **70**: 1063–1085.
- Farmer, C., and D. A. Hansell. 2007. Determination of dissolved organic carbon and total dissolved nitrogen in sea water. In A. G. Dickson, C. L. Sabine, and J. R. Christian [eds.], Guide to best practices for ocean CO<sub>2</sub> measurements - PICES Special Publication 3. North Pacific Marine Science Organization. Sidney, BC. Canada.
- Feng, Y., M. E. Warner, Y. Zhang, J. Sun, F. X. Fu, J. M. Rose, and D. A. Hutchins. 2008. Interactive effects of increased pCO<sub>2</sub>, temperature and irradiance on the marine coccolithophore *Emiliania huxleyi* (Prymnesiophyceae). Eur. J. Phycol. **43**: 87–98. doi:10.1080/09670260701664674
- Finkel, Z. V., J. Beardall, K. J. Flynn, A. Quigg, T. A. V. Rees, and J. A. Raven. 2010. Phytoplankton in a changing world: Cell size and elemental stoichiometry. J. Plankton Res. **32**: 119–137. doi:10.1093/plankt/fbp098
- Fogg, G. E. 1983. The ecological significance of extracellular products of phytoplankton photosynthesis. Botanica Marina **26**: 3–14. doi:10.1515/botm.1983.26.1.3
- Fu, F. X., M. E. Warner, Y. H. Zhang, Y. Y. Feng, and D. A. Hutchins. 2007. Effects of increased temperature and CO<sub>2</sub> on photosynthesis, growth, and elemental ratios in marine *Synechococcus* and *Prochlorococcus* (Cyanobacteria). J. Phycol. **43**: 485–496. doi:10.1111/j.1529-8817.2007.00355.x
- Fu, F. X., Y. H. Zhang, M. E. Warner, Y. Y. Feng, J. Sun, and D. A. Hutchins. 2008. A comparison of future increased CO<sub>2</sub> and temperature effects on sympatric *Heterosigma akashiwo* and *Prorocentrum minimum*. Harmful Algae **7**: 76–90. doi:10.1016/j.hal.2007.05.006
- Geider, R. J., J. La Roche, R. M. Greene, and M. Olaiola. 1993. Response of the photosynthetic apparatus of *Phaeodactylum tricornutum* (Bacillariophyceae) to nitrate, phosphate, or iron starvation. J. Phycol. **29**: 755–766. doi:10.1111/j.0022-3646.1993.00755.x
- Giordano, M., J. Beardall, and J. A. Raven. 2005. CO<sub>2</sub> concentrating mechanisms in algae: Mechanisms, environmental modulation, and evolution. Annu. Rev. Plant Biol. **56**: 99–131. doi:10.1146/annurev.arplant.56.032604.144052
- Goldman, J. C., D. A. Hansell, and M. R. Dennett. 1992. Chemical characterization of three large oceanic diatoms: Potential impact on water column chemistry. Mar. Ecol. Prog. Ser. **88**: 257–270. doi:10.3354/meps088257
- Goldman, J. C., J. J. McCarthy, and D. G. Peavey. 1979. Growth rate influence on the chemical composition of phytoplankton in oceanic waters. Nature **279**: 210–215. doi:10.1038/279210a0
- Grossart, H. P., M. Allgaier, U. Passow, and U. Riebesell. 2006. Testing the effect of CO<sub>2</sub> concentration on the dynamics of marine heterotrophic bacterioplankton. Limnol. Oceanogr. **51**: 1–11. doi:10.4319/lo.2006.51.1.0001
- Guillard, R. R. L., and J. H. Ryther. 1962. Studies of marine planktonic diatoms. I. *Cyclotella nana* Hustdt, and *Detonula confervacea* (Cleve) Gran. Can. J. Microbiol. **8**: 229–239. doi:10.1139/m62-029
- Hansell, D. A. 2005. Dissolved organic carbon reference material program. Eos Trans. Am. Geophys. Union **86**: 318–318. doi:10.1029/2005EO350003
- Hansell, D. A., and C. A. Carlson. 1998. Net community production of dissolved organic carbon. Glob. Biogeochem. Cycle **12**: 443–453. doi:10.1029/98GB01928
- Harrison, P. J., H. L. Conway, R. W. Holmes, and C. O. Davis. 1977. Marine diatoms grown in chemostats under silicate or ammonium limitation. 3. Cellular chemical composition and morphology of *Chaetoceros debilis*, *Skeletonema costatum*, and *Thalassiosira gravida*. Mar. Biol. **43**: 19–31. doi:10.1007/BF00392568
- Hopkinson, B. M., C. L. Dupont, A. E. Allen, and F. M. M. Morel. 2011. Efficiency of the CO<sub>2</sub>-concentrating mechanism of diatoms. Proc. Natl. Acad. Sci. USA **108**: 3830–3837. doi:10.1073/pnas.1018062108
- Hutchins, D. A., M. R. Mulholland, and F. X. Fu. 2009. Nutrient cycles and marine microbes in a CO<sub>2</sub>-enriched ocean. Oceanography **22**: 128–145. doi:10.5670/oceanog.2009.103
- Iglesias-Rodriguez, M. D., and others. 2008. Phytoplankton calcification in a high-CO<sub>2</sub> world. Science **320**: 336–340. doi:10.1126/science.1154122
- IPCC. 2007. Climate change 2007: The physical science basis. Contribution of Working Group I to the fourth assessment report of the intergovernmental panel on climate change, 996 p. Cambridge Univ. Press.

- Kester, D. R., I. W. Duedall, D. N. Connors, and Pytkowic, R. M. 1967. Preparation of artificial seawater. *Limnol. Oceanogr.* **12**: 176–179. doi: [10.4319/lo.1967.12.1.0176](https://doi.org/10.4319/lo.1967.12.1.0176)
- Kirchman, D. L., S. Y. Newell, and R. E. Hodson. 1986. Incorporation versus biosynthesis of leucine: Implications for measuring rates of protein synthesis and biomass production by bacteria in marine systems. *Mar. Ecol. Prog. Ser.* **32**: 47–59. doi:[10.3354/meps032047](https://doi.org/10.3354/meps032047)
- Koeve, W. 2004. Spring bloom carbon to nitrogen ratio of net community production in the temperate N. Atlantic. *Deep Sea Res. Part I Oceanogr. Res. Pap.* **51**: 1579–1600. doi:[10.1016/j.dsr.2004.07.002](https://doi.org/10.1016/j.dsr.2004.07.002)
- Körtzinger, A., W. Koeve, P. Kahler, and L. Mintrop. 2001. C : N ratios in the mixed layer during the productive season in the northeast Atlantic Ocean. *Deep Sea Res. Part I Oceanogr. Res. Pap.* **48**: 661–688. doi:[10.1016/s0967-0637\(00\)00051-0](https://doi.org/10.1016/s0967-0637(00)00051-0)
- Lampitt, R. S. 1985. Evidence for the seasonal deposition of detritus to the deep-sea floor and its subsequent resuspension. *Deep Sea Res. Part a Oceanogr. Res. Pap.* **32**: 885–897. doi: [10.1016/0198-0149\(85\)90034-2](https://doi.org/10.1016/0198-0149(85)90034-2)
- Langer, G., M. Geisen, K.-H. Baumann, J. Kläs, U. Riebesell, S. Thoms, and J. R. Young. 2006. Species-specific responses of calcifying algae to changing seawater carbonate chemistry. *Geochem. Geophys. Geosyst.* **7**: Q09006. doi:[10.1029/2005gc001227](https://doi.org/10.1029/2005gc001227)
- Leblanc, K., and others. 2012. A global diatom database - Abundance, biovolume and biomass in the world ocean. *Earth Syst. Sci. Data* **4**: 149–165. doi:[10.5194/essd-4-149-2012](https://doi.org/10.5194/essd-4-149-2012)
- Lewis, E., and D. W. R. Wallace. 1998. Program developed for CO<sub>2</sub> system calculations. ORNL/CDIAC-105. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S. Department of Energy.
- Logan, B. E., U. Passow, A. L. Alldredge, H. P. Grossart, and M. Simon. 1995. Rapid formation and sedimentation of large aggregates is predictable from coagulation rates (half-lives) of transparent exopolymer particles (TEP). *Deep Sea Res. Part II Top. Stud. Oceanogr.* **42**: 203–214. doi:[10.1016/0967-0645\(95\)00012-f](https://doi.org/10.1016/0967-0645(95)00012-f)
- Mari, X., S. Beauvais, R. Lemee, and M. L. Pedrotti. 2001. Non-redfield C : N ratio of transparent exopolymeric particles in the northwestern Mediterranean Sea. *Limnol. Oceanogr.* **46**: 1831–1836. doi:[10.4319/lo.2001.46.7.1831](https://doi.org/10.4319/lo.2001.46.7.1831)
- McGinnis, K. M., T. A. Dempster, and M. R. Sommerfeld. 1997. Characterization of the growth and lipid content of the diatom *Chaetoceros muelleri*. *J. Appl. Phycol.* **9**: 19–24. doi:[10.1023/a:1007972214462](https://doi.org/10.1023/a:1007972214462)
- Moran, X. A. G., A. Lopez-Urrutia, A. Calvo-Díaz, and W. K. W. Li. 2010. Increasing importance of small phytoplankton in a warmer ocean. *Glob. Change Biol.* **16**: 1137–1144. doi:[10.1111/j.1365-2486.2009.01960.x](https://doi.org/10.1111/j.1365-2486.2009.01960.x)
- Nelson, D. M., P. Treguer, M. A. Brzezinski, A. Leynaert, and B. Queguiner. 1995. Production and dissolution of biogenic silica in the ocean. Revised global estimates, comparison with regional data and relationship to biogenic sedimentation. *Glob. Biogeochem. Cycle* **9**: 359–372. doi: [10.1029/95gb01070](https://doi.org/10.1029/95gb01070)
- Passow, U. 2000. Formation of transparent exopolymer particles, TEP, from dissolved precursor material. *Mar. Ecol. Prog. Ser.* **192**: 1–11. doi:[10.3354/meps192001](https://doi.org/10.3354/meps192001)
- Passow, U. 2002. Transparent exopolymer particles (TEP) in aquatic environments. *Prog. Oceanogr.* **55**: 287–333. doi: [10.1016/S0079-6611\(02\)00138-6](https://doi.org/10.1016/S0079-6611(02)00138-6)
- Passow, U., and A. L. Alldredge. 1995. A dye-binding assay for the spectrophotometric measurement of transparent exopolymer particles (TEP). *Limnol. Oceanogr.* **40**: 1326–1335. doi:[10.4319/lo.1995.40.7.1326](https://doi.org/10.4319/lo.1995.40.7.1326)
- Passow, U., and C. A. Carlson. 2012. The biological pump in a high CO<sub>2</sub> world. *Mar. Ecol. Prog. Ser.* **470**: 249–271. doi:[10.3354/meps09985](https://doi.org/10.3354/meps09985)
- Raven, J. A., and R. J. Geider. 1988. Temperature and algal growth. *New Phytol.* **110**: 441–461. doi:[10.1111/j.1469-8137.1988.tb00282.x](https://doi.org/10.1111/j.1469-8137.1988.tb00282.x)
- Reinfelder, J. R. 2011. Carbon concentrating mechanisms in eukaryotic marine phytoplankton, p. 291–315. In C. A. Carlson and S. J. Giovannoni [eds.], *Annual review of marine science*, v. 3.
- Reinfelder, J. R., A. M. L. Kraepiel, and F. M. M. Morel. 2000. Unicellular C4 photosynthesis in a marine diatom. *Nature* **407**: 996–999. doi:[10.1038/35039612](https://doi.org/10.1038/35039612)
- Riebesell, U., A. Kortzinger, and A. Oschlies. 2009. Sensitivities of marine carbon fluxes to ocean change. *Proc. Natl. Acad. Sci. USA* **106**: 20602–20609. doi:[10.1073/pnas.0813291106](https://doi.org/10.1073/pnas.0813291106)
- Riebesell, U., D. A. Wolf-Gladrow, and V. Smetacek. 1993. Carbon dioxide limitation of marine phytoplankton growth rates. *Nature* **361**: 249–251. doi:[10.1038/361249a0](https://doi.org/10.1038/361249a0)
- Riebesell, U., I. Zondervan, B. Rost, P. D. Tortell, R. E. Zeebe, and F. M. M. Morel. 2000. Reduced calcification of marine plankton in response to increased atmospheric CO<sub>2</sub>. *Nature* **407**: 364–367. doi:[10.1038/35030078](https://doi.org/10.1038/35030078)
- Riebesell, U., and others. 2007. Enhanced biological carbon consumption in a high CO<sub>2</sub> ocean. *Nature* **450**: 545–548. doi:[10.1038/Nature06267](https://doi.org/10.1038/Nature06267)
- Roberts, K., E. Granum, R. C. Leegood, and J. A. Raven. 2007. C3 and C4 pathways of photosynthetic carbon assimilation in marine diatoms are under genetic, not environmental, control. *Plant Physiol.* **145**: 230–235. doi: [10.1104/pp.107.102616](https://doi.org/10.1104/pp.107.102616)
- Rost, B., U. Riebesell, S. Burkhardt, and D. Sultemeyer. 2003. Carbon acquisition of bloom-forming marine phytoplankton. *Limnol. Oceanogr.* **48**: 55–67. doi:[10.4319/lo.2003.48.1.0055](https://doi.org/10.4319/lo.2003.48.1.0055)
- Rost, B., U. Riebesell, and D. Sultemeyer. 2006. Carbon acquisition of marine phytoplankton: Effect of photoperiod length. *Limnol. Oceanogr.* **51**: 12–20. doi:[10.4319/lo.2006.51.1.0012](https://doi.org/10.4319/lo.2006.51.1.0012)



- Rost, B., I. Zondervan, and D. Wolf-Gladrow. 2008. Sensitivity of phytoplankton to future changes in ocean carbonate chemistry: Current knowledge, contradictions and research directions. *Mar. Ecol. Prog. Ser.* **373**: 227–237. doi:10.3354/meps07776
- Sambrotto, R. N., and others. 1993. Elevated consumption of carbon relative to nitrogen in the surface ocean. *Nature* **363**: 248–250. doi:10.1038/363248a0
- Sarthou, G., K. R. Timmermans, S. Blain, and P. Treguer. 2005. Growth physiology and fate of diatoms in the ocean: A review. *J. Sea Res.* **53**: 25–42. doi:10.1016/j.seares.2004.01.007
- Simon, M., and F. Azam. 1989. Protein content and protein synthesis rates of planktonic marine bacteria. *Mar. Ecol. Prog. Ser.* **51**: 201–213. doi:10.3354/meps051201
- Smith, D. C., and F. Azam. 1992. A simple, economical method for measuring bacterial protein synthesis rates in seawater using <sup>3</sup>H-leucine. *Mar. Microb. Food Webs* **6**: 107–114.
- Sommer, U., and K. Lengfellner. 2008. Climate change and the timing, magnitude, and composition of the phytoplankton spring bloom. *Glob. Change Biol.* **14**: 1199–1208. doi:10.1111/j.1365-2486.2008.01571.x
- Sundquist, E. T., L. N. Plummer, and T. M. L. Wigley. 1979. Carbon dioxide in the ocean surface: The homogeneous buffer factor. *Science* **204**: 1203–1205. doi:10.1126/science.204.4398.1203
- Taucher, J., K. G. Schulz, T. Dittmar, U. Sommer, A. Oschlies, and U. Riebesell. 2012. Enhanced carbon overconsumption in response to increasing temperatures during a mesocosm experiment. *Biogeosciences* **9**: 3531–3545. doi:10.5194/bg-9-3531-2012
- Toggweiler, J. R. 1993. Carbon overconsumption. *Nature* **363**: 210–211. doi:10.1038/363210a0
- Tortell, P. D., and others. 2008. CO<sub>2</sub> sensitivity of Southern Ocean phytoplankton. *Geophys. Res. Lett.* **35**: L04605. doi:10.1029/2007gl032583
- Treguer, P., D. M. Nelson, A. J. Vanbennekom, D. J. Demaster, A. Leynaert, and B. Queguiner. 1995. The silica balance in the world ocean: A reestimate. *Science* **268**: 375–379. doi:10.1126/science.268.5209.375
- Trimborn, S., D. Wolf-Gladrow, K. U. Richter, and B. Rost. 2009. The effect of pCO<sub>2</sub> on carbon acquisition and intracellular assimilation in four marine diatoms. *J. Exp. Mar. Biol. Ecol.* **376**: 26–36. doi:10.1016/j.jembe.2009.05.017
- Wetz, M. S., and P. A. Wheeler. 2003. Production and partitioning of organic matter during simulated phytoplankton blooms. *Limnol. Oceanogr.* **48**: 1808–1817. doi:10.4319/lo.2003.48.5.1808
- Wetz, M. S., and P. A. Wheeler. 2007. Release of dissolved organic matter by coastal diatoms. *Limnol. Oceanogr.* **52**: 798–807. doi:10.4319/lo.2007.52.2.0798
- Wohlers, J., and others. 2009. Changes in biogenic carbon flow in response to sea surface warming. *Proc. Natl. Acad. Sci. USA* **106**: 7067–7072. doi:10.1073/pnas.0812743106
- Wolf-Gladrow, D., and U. Riebesell. 1997. Diffusion and reactions in the vicinity of plankton: A refined model for inorganic carbon transport. *Mar. Chem.* **59**: 17–34. doi:10.1016/s0304-4203(97)00069-8
- Wu, Y., D. A. Campbell, A. J. Irwin, D. J. Suggett, and Z. Finkel. 2014. Ocean acidification enhances the growth rate of larger diatoms. *Limnol. Oceanogr.* **59**: 1027–1034. doi:10.4319/lo.2014.59.3.1027
- Zeebe, R. E., and D. Wolf-Gladrow. 2001. CO<sub>2</sub> in seawater: Equilibrium, kinetics, isotopes. Elsevier.

#### Acknowledgments

We thank Julia Sweet for assistance with pH and TEP measurements, as well as Maverick Carey and Elisa Halewood for technical assistance with DOC and TDN measurements. We would also like to thank two anonymous reviewers and the editorial office for helpful and constructive comments on the manuscript. This study was supported by the German Research Foundation (Deutsche Forschungsgemeinschaft, DFG), the German Academic Exchange Service (DAAD), and the National Science Foundation (NSF; OCE-1041038).

Submitted 31 October 2014

Revised 16 January 2015

Accepted 13 January 2015

Associate editor: John Albert Raven